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Development and testing of bioelectrochemical reactors converting wastewater organics into hydrogen peroxide

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Abstract

In a bioelectrochemical system, the energy content in dissolved organic matter can be used to power the production of hydrogen peroxide (H_2O_2), which is a potentially useful chemical at wastewater treatment plants. H_2O_2 can be produced by the cathodic reduction of oxygen. We investigated four types of gas-diffusion electrodes (GDEs) for this purpose. A GDE made of carbon nanoparticles bound with 30% polytetrafluoroethylene (PTFE) (wt./wt.C) to a carbon fiber paper performed best and catalyzed H_2O_2 production from oxygen in air with a coulombic efficiency of 95.1%. We coupled the GDE to biological anodes in two bioelectrochemical reactors. When the anodes were fed with synthetic wastewater containing acetate they generated a current of up to ~ 0.4 mA/mL total anode compartment volume. H_2O_2 concentrations of $\sim 0.2\%$ and $\sim 0.5\%$ could be produced in 5 mL catholyte in 9 hrs and 21 hrs, respectively. When the anodes were fed with real wastewater, the generated current was ~ 0.1 mA/mL and only 84 mg/L of H_2O_2 was produced.

Keywords: Bioelectrochemical systems; hydrogen peroxide; wastewater treatment; microbial fuel cell

INTRODUCTION

In bioelectrochemical systems (BESs), organics are oxidized by bacteria and the electrons are shuttled into an electrode called the anode. The electrons flow as a current through an external circuit to another electrode, the cathode, where a compound is reduced (Rozendal et al., 2008). The most commonly investigated type of BES is the microbial fuel cell (MFC), which is used to recover electrical energy from organic compounds in e.g. wastewater. In a MFC, oxygen is typically reduced to water at the cathode. Electrical energy can be recovered because the overall reaction, i.e. oxidation of organic compounds at the anode and oxygen reduction at the cathode is thermodynamically favorable (Logan et al., 2006). Another type of BES is the microbial electrolysis cell (MEC) for hydrogen production. Protons are reduced to hydrogen gas at the cathode. This occurs at a redox potential that is lower than the oxidation of organics at the anode, which means that the overall reaction is thermodynamically unfavorable. Thus, a small voltage input is needed to drive the reactions. However, because of the oxidation of organics at the anode, the energy value of the produced hydrogen can be significantly higher than the required input of electrical energy (Liu et al., 2005; Rozendal et al., 2006). In addition to producing electrical energy and hydrogen, BESs could potentially be used for a variety of other purposes. Examples include methane production

(Cheng et al., 2009), denitrification (Clauwaert et al., 2007), caustic production (Rabaey et al., 2010) or redistribution of alkalinity (Modin et al., 2011), ethanol production (Steinbusch et al., 2010), and perchlorate reduction (Trash et al., 2007).

We have focused on using a BES for production of hydrogen peroxide (H_2O_2), which is a chemical that is useful in many applications. In a wastewater treatment plant, H_2O_2 could be used for effluent disinfection, advanced oxidation of recalcitrant organics, and cleaning of membranes in membrane bioreactor plants. Currently, most of the world's H_2O_2 is produced by the reduction of oxygen using the anthraquinone oxidation process, which requires use of hydrogen and non-aqueous solvents (Pletcher, 1999). However, H_2O_2 can also be produced electrochemically by the cathodic reduction of oxygen. Rozendal et al. (2009) showed that a BES can be used to produce H_2O_2 with a low energy cost. They produced a concentration of 0.13% with a voltage input of 0.5 V (Rozendal et al., 2009). Fu et al. (2010) produced H_2O_2 in a MFC, but could only generate a concentration of around 80 mg/L (Fu et al., 2010). MFCs have also been investigated for generation of Fenton's reagent at the cathode (Zhuang et al., 2010; Feng et al., 2010).

In this study we developed and tested bioelectrochemical reactors for conversion of organic matter into H_2O_2 . We assumed a concentration of 0.2 to 0.5% would be sufficient to apply H_2O_2 as a cleaning agent in a membrane bioreactor treatment plant (Grelot et al., 2008); thus, producing a concentration in this range was our target. First, a gas-diffusion cathode was developed that allowed the production of sufficiently high concentrations of H_2O_2 . Gas-diffusion electrodes (GDEs) are energy-efficient as they allow passive diffusion of oxygen from the air instead of air-sparging of the liquid. Second, the cathode was combined with a biological anode fed with synthetic wastewater containing acetate. The operational conditions of the biological anode were optimized to allow high current production. Third, the reactors were tested with real wastewater.

METHODS

Development of a gas-diffusion cathode

The GDEs were constructed by painting a catalyst onto a hydrophobic carbon fiber paper (Toray TGP-H-090). Four conditions were tested: (1) the carbon fiber paper without any catalyst, (2) the paper coated with carbon black (Cabot Black Pearls 2000) and 480% PTFE (wt. PTFE/wt. C), (3) the paper coated with 200 mesh graphite powder (Alfa Aesar) and 480% PTFE, and (4) the paper coated with carbon black and 30% PTFE. The GDEs were prepared by painting a slurry containing carbon and PTFE onto the carbon fiber paper, allowing the paper to air dry and then sintering it for about 20 min at 350°C. PTFE is used as a binder that glues the catalysts layer to the carbon fiber paper. The performance of the GDEs was tested in an electrochemical cell consisting of 2.7 ml anode and cathode compartments separated by a Nafion 117 proton exchange membrane. The cathode surface area was 1.3 cm² and 50 ml of 50 mM NaCl was circulated through the cathode compartment using a peristaltic pump. The best performing GDE was then again tested in a slightly larger reactor having a 3.1 cm² cathode surface area and 12 ml cathode volume.

Bioelectrochemical reactor

Two cylindrical (diameter = 2 cm) bioelectrochemical reactors were constructed. The reactors contained GDEs and Nafion membranes with surface areas of 3.14 cm². The width of the anode compartments were 3 cm resulting in total volumes of approximately 9.4 ml. A graphite rod (diameter=0.615cm, length=3cm, Alfa Aesar) alone or together with a carbon fiber felt (4x3x0.5cm, TMIL) was used as anode. In Reactor A, the width of the cathode compartment was 3 cm whereas in Reactor B, it was 1 cm. The anode was fed with either a nutrient solution containing acetate

(Modin et al., 2010) or real wastewater that had passed the preliminary treatment steps in a wastewater treatment plant in Tokyo. The catholyte was always 50 mM NaCl.

Analytical methods

Cyclic voltammetry was performed using a potentiostat (KP07, Bank IC) and a data acquisition device (USB-6211, National Instruments) controlled with LabView software. The cathode was scanned at 10 mV/s whereas the anode was scanned at 1 mV/s. The anode and cathode potentials were measured and controlled against Ag/AgCl reference electrodes (BAS Inc.) but are reported against the Normal Hydrogen Electrode (NHE) assuming an offset of 0.197 V. H₂O₂ concentration was measured spectrophotometrically using the titanium oxalate method (Sellers, 1980). Acetate concentrations were measured using ion chromatography (Metrohm 761 Compact IC). Coulombic efficiency of cathodic H₂O₂ production was calculated using Equation 1.

$$CE = 100\% \times \frac{\Delta[H_2O_2] \times V \times (2/34)}{I \times (\Delta t / F)} \quad (1)$$

where *CE* is the coulombic efficiency (%), $\Delta[H_2O_2]$ is the change in concentration over time *t* (mg/L), *V* is the catholyte volume (L), 2/34 converts mg H₂O₂ into mmol e⁻ required to reduce O₂ to H₂O₂, *I* is the current (mC/s), Δt is length of time interval (s), *F* is Faraday's constant (96485.3 mC/mmol e⁻).

RESULTS AND DISCUSSION

Development of a gas-diffusion electrode

A comparison of the four tested GDEs is shown in Figure 1. The GDEs coated with carbon nanoparticles (#2 and #4) supplied higher currents than the GDE without catalyst or coated with graphite powder, probably because of a significantly larger surface area. GDE 4, i.e. carbon fiber paper coated with carbon black and 30% PTFE, performed the best (Fig. 1A). It supplied the largest cathodic current and it produced H₂O₂ with 95.1% efficiency when operated for 253 min at a constant cathode potential of -0.31 V vs NHE (Fig. 1B). GDE 4 probably generated a higher current than GDE 2 because of the lower content of PTFE (30% compared to 480%). PTFE is hydrophobic; thus, a lower PTFE fraction led to a larger portion of the carbon nanoparticle layer being exposed to water, increasing the total surface area available for reaction.

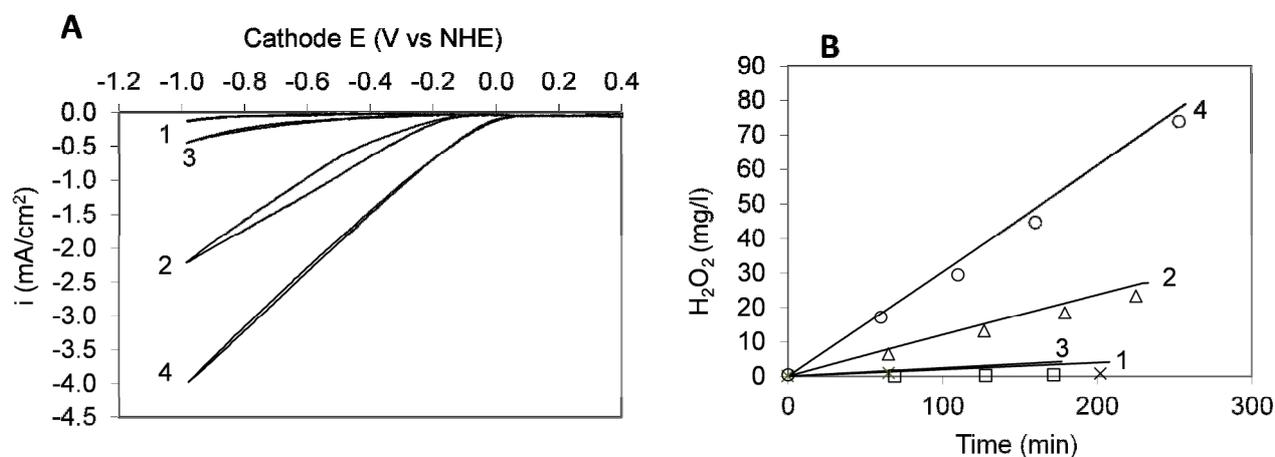


Figure 1. (A) Cyclic voltammograms of the four GDEs tested (cathodic current is negative). (B) Theoretical (line) and measured (symbols) H₂O₂ concentrations produced during operation of the GDEs at -0.31 V vs NHE. (1) Carbon fiber paper without any catalyst. (2) Carbon fiber paper

coated with carbon black and 480% PTFE. (3) Carbon fiber paper coated with 200 mesh graphite powder and 480% PTFE. (4) Carbon fiber paper coated with carbon black and 30% PTFE.

H₂O₂ production by GDE 4 was subsequently tested in three 9-h experiments with 12 ml catholyte volume at cathode potentials of -0.16, -0.31, and -0.46 V vs NHE. The resulting concentrations and coulombic efficiencies were 374 mg/L (66.4%), 1139 mg/L (78.6%), and 3200 mg/L (87.2%), respectively. The 3200 mg/L concentration was within the target range of 0.2-0.5%, so GDE 4 was used in the bioelectrochemical reactors.

Bioelectrochemical H₂O₂ production

Two GDEs of type 4 were made and used in the two bioelectrochemical reactors. The anodes in both reactors were initially single graphite rods. The reactors were inoculated with the effluent from a working microbial fuel cell. Initially both reactors were operated as microbial fuel cells with 1000 ohm resistors connected between the anode and cathode; this resistance was lowered to 100 ohm on day 8 and to 10 ohm on day 45. After 40 days of operation, a carbon felt was attached to each anode to increase the total surface area available for bacterial growth and thereby increase the current delivered by the anode. On day 50, we started recirculating the anolyte through the anode chambers at a flow rate of 50 mL/min. On day 86, real wastewater was fed to the anodes. The ability of the anodes to deliver current was evaluated using cyclic voltammetry (Figure 2). We can see that attaching a carbon felt to the anode increased current (compare day 37 and day 50 without recirculation) as this increased the surface area available for bacterial growth. During cyclic voltammetry, the anolyte was fed continuously to the anode chambers at a flow rate of 3.2 mL/min. Recirculating the anolyte at 50 mL/min improved current output (compare the voltammograms from day 50). Using real wastewater as feed reduced the current output compared to synthetic wastewater (compare day 97 and day 50 with recirculation).

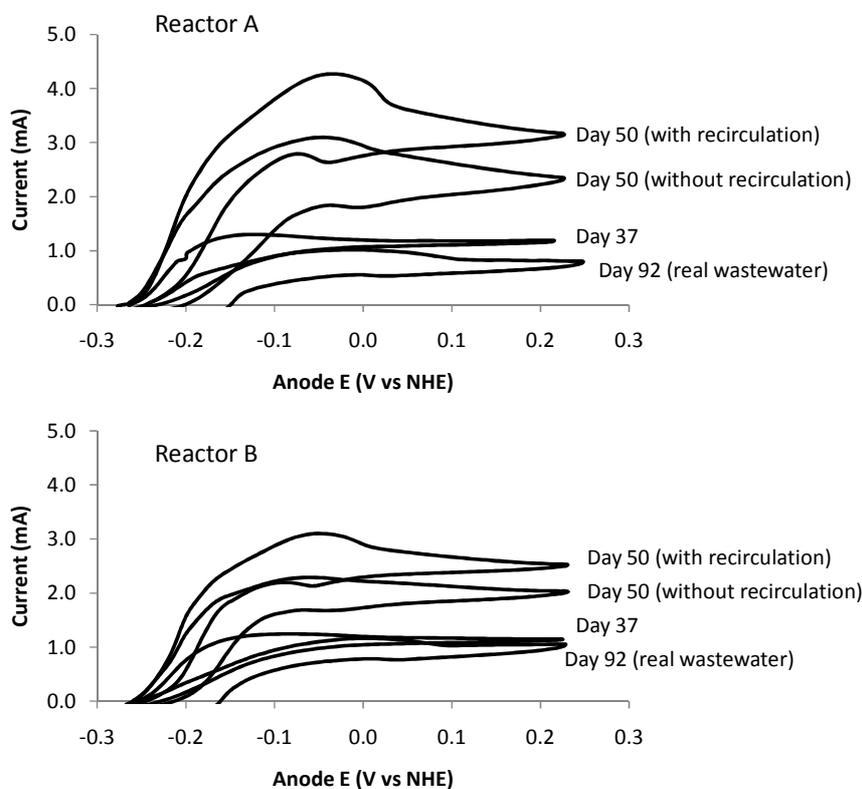


Figure 2. Cyclic voltammograms of the anode in Reactor A (upper) and Reactor B (lower). The current is plotted against the anode potential.

H₂O₂ production was investigated by loading the cathode chambers with known volumes of 50 mM NaCl solution and measuring the H₂O₂ concentration after a fixed time interval. The anode potential was controlled at -0.11 V vs NHE and fed continuously with anolyte at a flow rate of 1.6 mL/min. The results are summarized in Table 1.

On day 40, 47, and 51, 2-h tests were carried out with both reactors. Higher concentrations were produced in Reactor B since it had smaller catholyte volume. Higher current led to higher H₂O₂ concentration as well as higher coulombic efficiency (CE). On day 65, a 9-h test was carried out with Reactor B with the aim of reaching the 0.2-0.5% target concentration. This was successful as over 0.2% could be produced with an energy input of 1.01 kWh/kgH₂O₂ and a CE of 61%. The required energy input was low compared to abiotic electrochemical reactors for H₂O₂ production, which may require 4.4-8.9 kWh/kgH₂O₂ (Foller and Bombard, 1995).

On day 66, we operated the reactors as MFCs with a 10 ohm resistor connected between anode and cathode. After 72.1 h operation, the H₂O₂ concentrations were approximately 0.06% and 0.08% in A and B, respectively. This means that the reactors could produce H₂O₂ and electrical energy simultaneously.

Concentrations of nearly 0.5% H₂O₂ could be produced after 46.9 h in Reactor A and 21 h in Reactor B. On day 104, another 21-h test was conducted with Reactor B using real wastewater. However, only 83.5 mg/L was produced, the CE was low, and the energy input was high.

There appears to be a correlation between final catholyte pH and CE. When the pH was over 11.28, the CE was over 50.3%. When the pH was lower than 10.79, the CE was lower than 28.1%. A reason for low CE may be cathodic reduction of H₂O₂ to H₂O. The pK_a of H₂O₂ is 11.62. If the pH is near or over this pH value, a large portion of the H₂O₂ will be present as the anion HO₂⁻. Being negatively charged, HO₂⁻ will be repelled from the cathode, which is also negatively charged (Qiang et al., 2002). This would prevent further reduction to H₂O and may explain the improved CE at higher pH values. Another reason for low CE could be self-decomposition of H₂O₂ according to Equation 2. This self-decomposition may have been catalyzed by metal ions migrating from the anode compartment through the cation exchange membrane to the cathode compartment. Diffusion of H₂O₂ to the anode compartment could also have resulted in lowering the CE.



Low current was also correlated with low CE, which is especially evident in the tests with the reactors operated as MFCs and in the test with real wastewater, which both had the lowest currents and the lowest CE. Low current corresponds to a low H₂O₂ production rate, which means that self-decomposition processes may have been more dominating. Low current also means that the pH rise in the cathode will to a higher degree be counteracted by dissolution of CO₂ from air. Thus, to improve the CE, a bioelectrochemical reactor should be designed with high anode surface area, which would maximize the generated current in relation to the cathode volume.

Table 1. H₂O₂ production tests. Reactor A had a catholyte volume of 11 mL and Reactor B had 5 mL. The anode was controlled at a potential of -0.11 V vs NHE except on day 66 when the reactors were operated as MFCs.

Reactor A	Day 40	Day 47	Day 51		Day 66	Day 76	
Run time (h)	2	2	2		72.1	46.9	
H ₂ O ₂ conc. (mg/L)	23.1	160.0	201.3		601.9	4947.5	
Final catholyte pH	10.54	11.4	11.28		10.54	12.35	
CE ^a	18.4%	50.3%	64.9%		13.5%	51.3%	
Energy use (kWh/kgH ₂ O ₂)	1.17	2.29	1.63		-238.2 ^b	2.27	
Average I (mA)	1.08	2.75	2.69		0.91	3.47	
Reactor B	Day 40	Day 47	Day 51	Day 65	Day 66	Day 69	Day 104
Run time (h)	2	2	2	9	72.1	21	21
H ₂ O ₂ conc. (mg/L)	76.9	357.8	643.0	2283.9	809.1	4588.8	83.5
Final catholyte pH	10.55	11.28	11.60	11.67	10.79	11.97	10.3
CE ^a	28.1%	65.0%	75.8%	61.0%	7.3%	59.3%	4.8%
Energy use (kWh/kgH ₂ O ₂)	0.30	0.75	1.32	1.01	-418.4 ^b	1.77	18.2
Average I (mA)	1.08	2.17	3.34	3.28	0.95	2.90	0.65

^aCoulombic Efficiency (percentage of current charge equivalents used for H₂O₂ production).

^bNegative values means that power was produced instead of being consumed.

Characterization of bioelectrochemical reactors

The reactors were characterized by stepwise increasing the current and monitoring the resulting anode potential, cathode potential, and cell voltage. The cathode compartment of Reactor B was too small to accommodate a reference electrode; therefore, results are only shown for Reactor A. Figure 3 shows results from experiments carried out with synthetic wastewater on 86 and with real wastewater on day 99. The resistances associated with the anode reaction, cathode reaction, and the internal conductivity of the cell were estimated based on the slopes of the lines. The total resistance increased from 535±20Ω on day 86 to 1057±70Ω on day 99. As seen in Fig. 3, the cathode performance remained largely unchanged, so the deterioration in performance was caused by worse anode performance and higher internal ohmic resistance. The worse anode performance can be attributed to the bioanode culture being acclimated to acetate and the real wastewater not containing as much of this substance as the synthetic wastewater. The ohmic resistance increased from 342±29Ω to 539±103Ω, which can be explained by the lower conductivity of the real wastewater, 127 mS/m compared to 777 mS/m for the synthetic wastewater.

With synthetic wastewater, the anode potential was lower than the cathode potential up to a current of about 2.3 mA. However, because of the ohmic resistance of the cell, a voltage input was required for currents exceeding 0.6 mA. Similarly for real wastewater, the anode potential was lower than the cathode potential up to 0.7 mA, but a voltage input was required over 0.3 mA. This indicates that if the ohmic resistance of the cell could be minimized, high concentrations of H₂O₂ could potentially be produced without energy input.

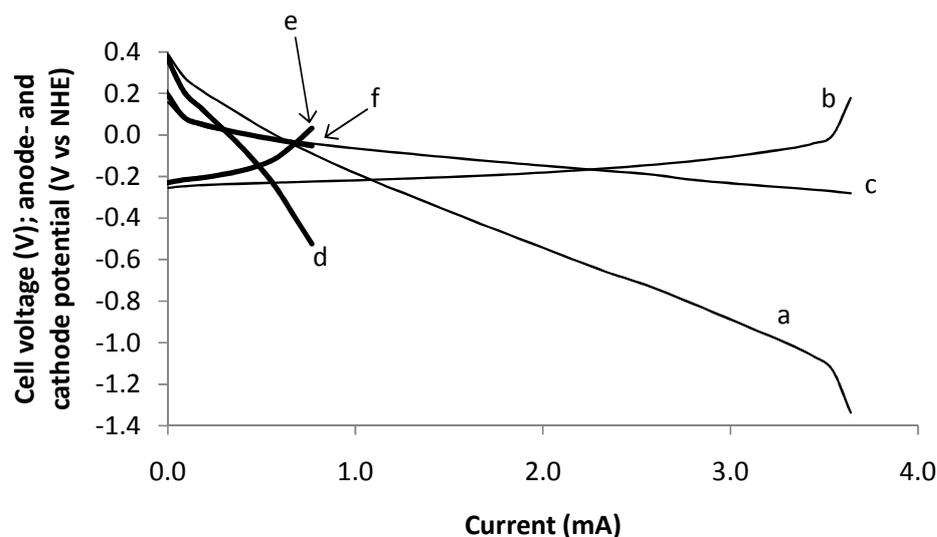


Figure 3. Cell voltage (a,d), anode potential (b,e), and cathode potential (c,f) on day 86 (a,b,c, thin lines, synthetic wastewater) and day 99 (d,e,f, thick lines, real wastewater).

The cation exchange membrane and GDEs were exchanged in the reactors on day 59. A white layer could be observed on the cathode side of the membrane (Figure 4). This was probably calcium carbonate deposits caused by calcium ions migrating from the anode compartment combining with carbonate ions present at high pH in the catholyte. This may have contributed to the high ohmic resistances of the cells. The GDEs seemed unaffected by these deposits.

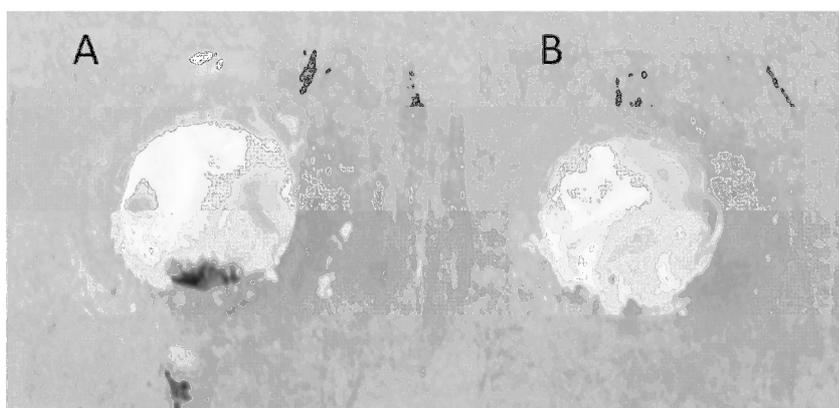


Figure 4. Nafion membranes harvested on day 59 of the experiment. White deposits were found on the cathode sides. The membranes are normally clear.

CONCLUSIONS

We made four GDEs and found that carbon nanoparticles bound with 30% PTFE could catalyze oxygen reduction to H_2O_2 with 95.1% coulombic efficiency. A bioelectrochemical reactor constructed with this GDE and fed with synthetic wastewater to the anode could produce over 0.2% H_2O_2 in 9h or 0.5% in 21h. These are concentrations that would likely be practically useful for membrane cleaning in MBR treatment plants.

Fed with real wastewater, the performance of the reactor deteriorated. The bioanode could only deliver a current less than a third in magnitude of the current delivered with synthetic wastewater. Furthermore, the internal resistance of the reactor increased because of the lower conductivity of the

real wastewater. Another problem with the reactor was white deposits (probably calcium carbonate) on the cation exchange membrane.

The study showed that bioelectrochemical systems can be used to produce practically relevant concentrations of H₂O₂ with a low input of electrical energy. However, the bioanode must be designed to deliver current of sufficiently high magnitude. In our experiment, high H₂O₂ concentrations (>0.2%) were produced with currents over 0.32 mA/mL cathode volume. A volume of 0.006 L of 0.2% H₂O₂ was produced per L synthetic wastewater fed to the anode chamber with an electric power consumption of 1.01 kWh/kgH₂O₂.

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REFERENCES

- Cheng, S., Xing, D., Call, D.F., Logan, B.E. (2009). Direct biological conversion of electrical current into methane by electromethanogenesis. *Environmental Science & Technology*, **43**, 3953-3958.
- Clauwaert, P., Rabaey, K., Aelterman, P., de Schamphelaire, L., Pham, T.H., Boeckx, P., Boon, N., Verstraete, W. (2007). Biological denitrification in microbial fuel cells. *Environmental Science & Technology* **41**, 3354-3360.
- Feng, C.H., Li, F-B., Mai, H-J., Li, X-Z. (2010). Bio-electro-fenton process driven by microbial fuel cell for wastewater treatment. *Environmental Science & Technology*, **44**, 1875-1880.
- Foller, P.C., Bombard, R.T. (1995). Processes for the production of mixtures of caustic soda and hydrogen peroxide via the reduction of oxygen. *Journal of Applied Electrochemistry*, **25**, 613-627.
- Fu, L., You, S-J., Yang, F-L., Gao, M-M., Fang, X-H., Zhang, G-Q. (2010). Synthesis of hydrogen peroxide in microbial fuel cell. *Journal of Chemical Technology and Biotechnology*, **85**, 715-719.
- Grelot, A., Machinal, C., Drouet, K., Tazi-Pain, A., Schrotter, J.C., Grasmick, A., Grinwis, S. (2008). In search of alternative cleaning solutions for MBR plants. *Water Science and Technology*, **58(10)**, 2041.
- Liu, H., Grot, S., Logan, B.E. (2005). Electrochemically assisted microbial production of hydrogen from acetate. *Environmental Science & Technology*, **39**, 4317-4320.
- Logan, B.E., Hamelers, B., Rozendal, R., Schröder, U., Keller, J., Freguia, S., Aelterman, P., Verstraete, W., Rabaey, K. (2006). Microbial fuel cells: methodology and technology. *Environmental Science & Technology*, **40(17)**, 5181-5192.
- Modin, O., Fukushi, K., Rabaey, K., Rozendal, R.A., Yamamoto, K. (2010). Bioelectrochemical hydrogen peroxide production – an opportunity for sustainable mitigation of membrane bioreactor fouling. *Proceedings of the Water Environment Federation, Biofilms 2010*, **309-321** (DOI: <http://dx.doi.org/10.2175/193864710798208566>).
- Modin, O., Fukushi, K., Rabaey, K., Rozendal, R.A., Yamamoto, K. (2011). Redistribution of wastewater alkalinity with a microbial fuel cell to support nitrification of reject water. *Water Research*, **45(8)**, 2691-2699.
- Pletcher, D. (1999) Indirect oxidation using electrogenerated hydrogen peroxide. *Acta Chemica Scandinavica*, **53**, 745-750.

- Rabaey, K., Butzer, S., Brown, S., Keller, J., Rozendal, R.A.. (2010). High current generation coupled to caustic production using a lamellar bioelectrochemical system. *Environmental Science & Technology*, **44**, 4315-4321.
- Rozendal, R.A., Hamelers, H.V.M., Euverink, G.J.W., Metz, S.J., Buisman, C.J.N. (2006). Principles and perspective of hydrogen production through biocatalyzed electrolysis. *International Journal of Hydrogen Energy*, **31**, 1632-1640.
- Rozendal, R.A., Hamelers, H.V.M., Rabaey, K., Keller, J., Buisman, C.J.N. (2008). Towards practical implementation of bioelectrochemical wastewater treatment. *Trends in Biotechnology*, **26(8)**, 450-459.
- Rozendal, R.A., Leone, E., Keller, J., Rabaey, K. (2009) Efficient hydrogen peroxide generation from organic matter in a bioelectrochemical system. *Electrochemistry Communications*, **11(9)**, 1752-1755.
- Qiang, Z, Chang, J-H., Huang, C-P. (2002). Electrochemical generation of hydrogen peroxide from dissolved oxygen in acidic solutions. *Water Research*, **36**, 85-94.
- Sellers, R.M. (1980) Spectrophotometric determination of hydrogen peroxide using potassium titanium(IV) oxalate. *Analyst*, **105**, 950-954.
- Steinbusch, K.J.J., Hamelers, H.V.M., Schaap, J.D., Kampman, C., Buisman, C.J.N. (2010). Bioelectrochemical ethanol production through mediated acetate reduction by mixed cultures. *Environmental Science & Technology*, **44**, 513-517.
- Trash, J.C., van Trump, J.I., Weber, K.A., Miller, E., Achenbach, L.A., Coates, J.D. (2007). Electrochemical stimulation of microbial perchlorate reduction. *Environmental Science & Technology*, **41**, 1740-1746.
- Zhuang, L., Zhou, S., Li, Y., Liu, T., Huang, D. (2010). In-situ Fenton enhanced cathodic reaction for sustainable increased electricity generation in microbial fuel cells. *Journal of Power Sources*, **195**, 1379-1382.