

REVIEW

Mind the gap—deficits in our knowledge of aspects impacting the bioavailability of phytochemicals and their metabolites—a position paper focusing on carotenoids and polyphenols

Torsten Bohn¹, Gordon J. McDougall², Amparo Alegría³, Marie Alminger⁴, Eva Arrigoni⁵, Anna-Marja Aura⁶, Catarina Brito^{7,8}, Antonio Cilla³, Sedef N. El⁹, Sibel Karakaya⁹, Marie C. Martínez-Cuesta¹⁰ and Claudia N. Santos^{7,8}

¹ Environmental Research and Innovation Department, Luxembourg Institute of Science and Technology, Belvaux, Luxembourg

² The James Hutton Institute, Invergowrie, Dundee, United Kingdom

³ Nutrition and Food Science Area, Faculty of Pharmacy, University of Valencia, Av. Vicente Andrés Estellés s/n, Burjassot, Valencia, Spain

⁴ Department of Chemical and Biological Engineering, Chalmers University of Technology, Gothenburg, Sweden

⁵ Agroscope, Institute for Food Sciences (IFS), Wädenswil, Switzerland

⁶ VTT Technical Research Centre of Finland, Espoo, Finland

⁷ IBET, Instituto de Biologia Experimental e Tecnológica, Oeiras, Portugal

⁸ Instituto de Tecnologia Química e Biológica António Xavier, Universidade Nova de Lisboa, Oeiras, Portugal

⁹ Ege University, Engineering Faculty, Food Engineering Department, Izmir, Turkey

¹⁰ Instituto de Investigación en Ciencias de la Alimentación CIAL (CSIC-UAM), Madrid, Spain

Various secondary plant metabolites or phytochemicals, including polyphenols and carotenoids, have been associated with a variety of health benefits, such as reduced incidence of type 2 diabetes, cardiovascular diseases, and several types of cancer, most likely due to their involvement in ameliorating inflammation and oxidative stress. However, discrepancies exist between their putative effects when comparing observational and intervention studies, especially when using pure compounds. These discrepancies may in part be explained by differences in intake levels and their bioavailability. Prior to exerting their bioactivity, these compounds must be made bioavailable, and considerable differences may arise due to their matrix release, changes during digestion, uptake, metabolism, and biodistribution, even before considering dose- and host-related factors. Though many insights have been gained on factors affecting secondary plant metabolite bioavailability, many gaps still exist in our knowledge. In this position paper, we highlight several major gaps in our understanding of phytochemical bioavailability, including effects of food processing, changes during digestion, involvement of cellular transporters in influx/efflux through the gastrointestinal epithelium, changes during colonic fermentation, and their phase I and phase II metabolism following absorption.

Received: October 15, 2014

Revised: March 27, 2015

Accepted: April 29, 2015

Keywords:

Biotransformation / Food processing / Microbiota / Mixed diet / Transporters

1 Introduction

Phytochemicals comprise a diverse group of secondary plant compounds, including polyphenols, carotenoids, triterpenes, phytosterols, glucosinolates, and many more. These compounds have aroused increasing interest in the area of

Colour Online: See the article online to view Fig. 1 in colour.

Correspondence: Dr. Torsten Bohn

E-mail: torsten.bohn@gmx.ch

Abbreviations: CVD, cardiovascular diseases; MCTs, monocarboxylic acid transporters; MRP, multidrug resistance protein; NEPP, nonextractable polyphenols; NPC1L1, Niemann-Pick C1-Like 1; SGLT-1, sodium-glucose linked transporter 1; T2D, type 2 diabetes; TOF, time-of-flight

nutrition and food science, due to their potential health benefits. Several epidemiological studies have suggested that their consumption or tissue concentration is associated with reduced risk of developing certain chronic diseases, namely, type 2 diabetes (T2D), cardiovascular diseases (CVD), and some types of cancer. For example, meta-analyses have suggested that the consumption of flavonoids from fruits and vegetables improved flow-mediated dilation, a marker of atherosclerosis [1], and suggested positive health effects of consuming carotenoids with respect to type 2 diabetes [2].

However, though a few examples of intervention studies with isolated compounds, e.g. resveratrol, or curcumin [3, 4], producing positive health effects exist, micronutrients and phytochemicals may not act efficiently in isolation but together with many other compounds in the food matrix, leading to synergistic effects [5]. It is thus important to determine efficacy, safety and underlying mechanisms of these compounds, especially when taken in pharmaceutical doses/in combination with other drugs. For example, Prasain et al. [6] reviewed risks and benefits of dietary versus supplemented/isolated flavonoids, stating that flavonoids can be detrimental in some settings and therefore are not universally safe. Adverse effects were also found following the ingestion of beta-carotene supplements administered to CVD patients [7,8], and vitamin C and E as well as beta-carotene supplementation failed to show health benefits with respect to CVD [9]. Supplementing individual flavonoids and isoflavonoids has also been met with criticism, e.g. due to endocrine disrupting properties for higher doses [10]. The reasons for this discrepancy are not fully understood, but could involve “missing” synergistic effects with the food matrix, altered digestion and release of the compounds or changed degradation patterns, i.e. modified bioavailability (the fraction of a compound that is absorbed and can be used for physiological functions and/or storage). Indeed, many reviews on bioavailability and bioactivity of phytochemicals are available [11–13]. Phytochemicals may have multiple functions in the human body, and depending on their dose they can exert both beneficial and deleterious effects [14].

This position paper attempts to identify the aspects of digestion, release, absorption and metabolism of food phytochemicals which are only poorly understood (Table 1) [15]. Even prior to ingestion processing factors (food texture, e.g. heat, temperature, or pressure application [16]) can impinge on the bioaccessibility (fraction of a compound that is released from the matrix and potentially available for further uptake and absorption) of bioactives [17]. Similarly, the influence of consuming a mixed diet, i.e. “real” complex meals, on bioaccessibility and absorption is poorly comprehended, and biochemical and physico-chemical aspects such as viscosity and surface tension surely play a role [18]. Following ingestion, enzyme concentrations, pH, and time of digestion all play a role and influence release kinetics and degradation patterns [19, 20]. A factor that is also poorly understood is the nature and bioactivity of metabolites formed during digestion, as following their fate is challenging. However, the

bioavailability of some compounds and their metabolites may be higher than previously assumed, as shown, e.g. in isotope studies with anthocyanins [21]. Some lipophilic compounds, such as carotenoids or triterpenes, require micellization, as may certain polyphenol aglycones [22]. Following release, pathways of absorption, i.e. active versus passive or paracellular routes remain largely marginally understood. The same is true for influx and efflux transporters in the epithelium. For example, several polyphenols were shown to be considerably better absorbed in the presence of additional polyphenols, blocking efflux transporters (to the gut), which normally reduce the intracellular concentration of such “xenobiotics” [19]. On the other hand, transporters to the basolateral side are hardly understood. Finally, many native phytochemicals undergo considerable metabolism in the human body, e.g. deglycosylation and glucuronidation/sulfation for polyphenols in the gut, cleavage by beta-carotene oxygenase 1 and beta-carotene dioxygenase 2 for carotenoids [23], and many additional reactions may occur in other tissues such as in the liver or in the colon, where bacterial fermentation significantly alters the structure and profile and thus the potential bioactivity of many plant compounds that are not absorbed in the small intestine [24–26].

In this paper, we aim to highlight gaps that have received either limited attention or which are far from being understood, but which may play pivotal roles in the bioavailability of phytochemicals (Fig. 1). To allow for better focus, this review will concentrate on polyphenols and their metabolites as the most abundant water-soluble compounds, and carotenoids as the most abundant lipophilic secondary plant compounds. For a more comprehensive overview on bioavailability aspects of specific compound classes, the reader is referred to other articles [22, 27, 28].

2 Food processing and matrix effects

Bioaccessibility, and further bioavailability, of phytochemicals starts with their content and composition of the raw plant material and how it is processed. The food matrix and structure and type of processing can have both positive and negative effects [29]. Plant matrix disruption and cell cluster disintegration due to applied processing steps are the main prerequisites for phytochemical liberation and bioaccessibility, but they may also lead to oxidation and/or degradation, thus potentially counterbalancing each other.

To understand the release of bioactive compounds during digestion, it is also important to know where they are located in the tissue. Carotenoids are found either in the chloroplast membrane or in chromoplasts [30]. The different physical forms of carotenoids in plant chromoplasts (crystals in tomato, lipid-dissolved in papaya and liquid-crystalline in mango) have major impacts on their liberation efficiency from the food matrix [31]. Polyphenols are generally present in vacuoles and the apoplast of plant cells, in conjugated form with mono- and polysaccharides, and proteins.

Table 1. Summary of major gaps of knowledge around phytochemical bioavailability

Stage	Knowledge gap	Examples	Reference examples
Food matrix	Physical state/compartmentalization of phytochemicals	Crystallinity of carotenoids in chromoplasts versus chloroplasts, polyphenols bound to cell wall (NEPP)	[31, 107]
Food preparation	Effect of cutting, mashing, grinding, peeling, trimming	Enzyme activation (e.g. polyphenol oxidase, alliinase) Concentration differences in morphological parts	[42, 43]
Food processing	Effect of cooking (heat, temperature, time, blanching)	Enhanced carotenoid content with mild conditions but decreased with severe treatments	[16, 52, 53]
	Refining processes	Bleaching, deodorization: decreasing carotenoid content Milling enhancing polyphenol extractability through surface area	[40–42]
	Nonthermal processing (HPP and PEF)	Both positive (polyphenols) and contradictory effects (carotenoids)	[67, 69]
Food mixtures	Effect of mixed or real meals	Enhanced availability of polyphenols with presence of sugars, ascorbic acid, fat. Stabilizing effects of polyphenols on other phytochemicals?	[77, 81, 193]
Gastric digestion	Effect of pH, depolymerization of large polyphenols, binding effects	Hydrophobic interactions, hydrogen bonding of polyphenols to proteins	[19]
Small intestine	Micelle formation Role of uptake transporters	Transition from matrix to oil phase, micelle size, number, and stability St. John's Wort components increased P-gp function	[22, 83]
	Efflux transporters cell→gut lumen	Efflux transporters such as P-gp, BCRP blocked by several polyphenols, e.g. flavonoids	[124]
	Efflux transporters cell→basolateral	Effect of polyphenols on efflux transporters such as MRP3	[194–196]
	Transporters gut lumen→cell	SRB-1, CD36, NPC1L1, ABCG5/G8: affecting carotenoid uptake – influence of polyphenols	[19, 197, 198]
Colon	Phase I/II interactions Influence of microflora Colonic absorption	Piperine increased curcumin absorption Metabolite formation, absorption of cleavage products?	[146] [102, 172, 190]
	Metabolites and phase I/II products	Metabolite information limited by lack of standards	[160, 161]
	Biotransformation, phase I and phase II metabolism	Need to develop/increased availability of more physiological cell models (liver stem cells, co-culture cell models) to study metabolites	[125, 152–155]
Tissues	Interaction with transporters in certain tissues (blood–brain barrier, placenta, testis), or excretory organs (liver, kidney)	Role of MRPs, MCTs, CD36	[140, 184, 185]

ABCG5/G8 = ATP-binding cassette sub-family G, member 5/8; BCRP = Breast cancer resistance protein; CD36 = cluster of differentiation; HPP = high pressure processing; MCTs = monocarboxylic acid transporters; MRP = multidrug resistance protein; NEPP = non-extractable polyphenols; NPC1L1 = Niemann-Pick C1-Like 1; PEF = pulsed electric fields; P-gp = P-glycoprotein; SRB-1 = scavenger receptor class B member 1.

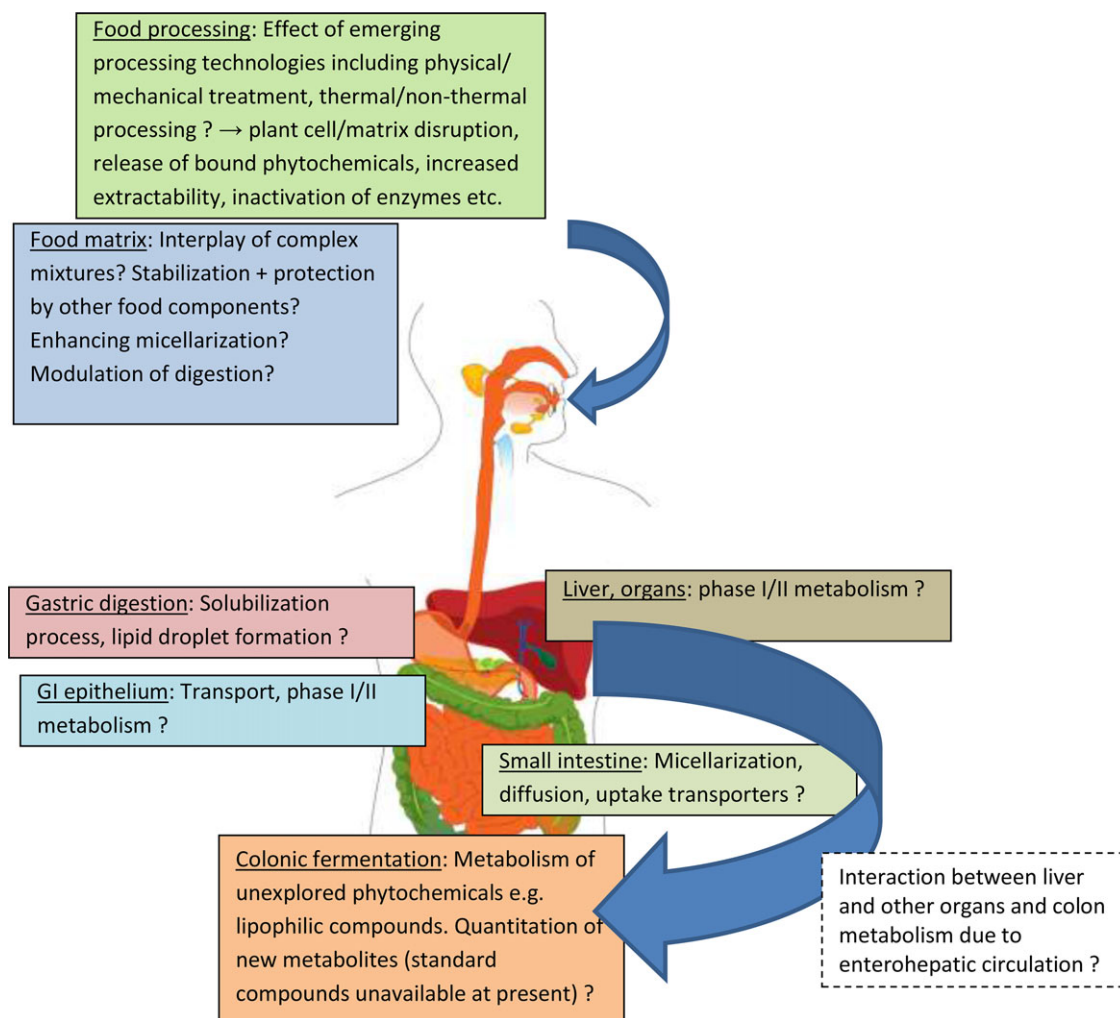


Figure 1. Overview of important stages during the digestion of food items and open questions related to phytochemical bioavailability.

Although studies have focused mainly on the cellular localization of carotenoids, polyphenols, and more rarely on glucosinolates, more research on their localization in different foods is needed. Little is known about other phytochemicals.

2.1 Effects of physical and mechanical treatments

Food preparation (such as grating, chopping, cutting, slicing, trimming, mashing, and juicing) may have great influence on the bioaccessibility of nutrients and bioactive compounds [30, 32]. The recovery of phytochemicals from intact, minimally processed fruits and vegetables can be different from juices as components must be released from the tissue matrix before being subject to degradation reactions, whereas those in the juices are subject to degradation from the offset. However, carotenoid bioaccessibility was greater from juiced than from raw or cooked puréed tissue, as chopping and homogenization disrupted the plant matrix [32]. The levels of carotenoids in sea buckthorn berries were lower if the berries

were extracted in water or juiced than after direct acetone extraction. This appears to be due to the complexation of carotenoids with pectins [33], and similar interactions may also occur between pectins and polyphenols, such as anthocyanins, during digestion [34].

Other phytochemicals are subject to endogenous enzymatic digestion. Chopping or crushing raw garlic releases alliinase which produces allicin, which breaks down to diallyl sulfides and then other metabolites, which may influence chronic diseases and certain cancers [35, 36]. Similarly, the glucosinolate glucobrassicin, which is found in cruciferous vegetables, is hydrolyzed when plant cells are damaged [37]. On the other hand, flavonoids seem relatively stable to mechanical processing [38]. Peeling and trimming steps may also influence phytochemical content. Trimming green leafy vegetables is known to be a main factor influencing carotenoid concentrations, since they are nearly exclusively present in leaves [39]. Also industrial refining processes, e.g. as applied to vegetable oils and cereals, may lead to high

losses in phytochemicals. Refining of palm oil decreased carotenoids by 99%, and polyphenols by 23% [40, 41].

Milling of cereals and grains has a great influence on phenolic acids [42]. It is well known that milling of the whole grain results in higher amounts of phenolics than conventional milling resulting in white flour [43]. Many studies have investigated the effects of refining on phytochemicals in edible oils and cereals but similar studies on vegetables and fruits (except juices) are largely absent.

In conclusion, although there are many studies reporting effects of food preparation on the bioaccessibility of polyphenols and carotenoids, understanding the mechanisms of chemical or enzymatic actions still requires extensive work. In addition, as novel methods are developed, further studies on how these processes impact bioaccessibility and bioavailability are warranted.

2.2 Effects of thermal and nonthermal treatments

Studies of thermal treatment on phytochemicals in foods have mainly focused on polyphenols, carotenoids and glucosinolates. Many reports indicate that thermal processing increases levels of free polyphenols [44–47], possibly due to the release of bound phenolics due to the breakdown of cell constituents, perhaps through releasing nonextractable polyphenols (NEPP). However, also polymerization/oxidation reactions may be responsible for apparent increases [44, 45], and the nature of these released polyphenols requires investigation. Similarly, carotenoid availability often increases after heat treatment [48–51], probably due to enhanced extractability following maceration of cells. However, severe heat treatment such as baking/sterilization of tomatoes [52, 53] or boiling of chili peppers [54] caused significant losses. Losses of glucosinolates in *Brassica* vegetables depended on processing time, type of vegetable, degree of cellular damage, and inactivation of myrosinase [55–60]. Overall, further research is required to better explain how thermal treatments impact bioaccessibility and bioavailability.

Nonthermal processing technologies have been revealed as useful tools to extend shelf-life and to preserve nutritional and functional characteristics of fruit and vegetable products [61]. However, there are scarce data on the effect of these emerging technologies on bioaccessibility and bioavailability of bioactive compounds. Considering bioaccessibility, studies have mainly focused on the effect of high pressure processing on carotenoids, with contradictory results. Some show improvements [16, 62–66], possibly related to the disintegration of cell clusters and disruption of cells containing carotenoids. Beneficial effects of a subsequent thermal treatment may weaken the physical barriers that enclose carotenoids and accelerate pectin degradation by β -elimination, resulting in cell wall softening. Decreased carotenoid bioaccessibility has been noted in some vegetables and fruits and may be due to firmer texture and/or trapping of carotenoids within fiber networks [62, 63]. Thus, processing variables (e.g. particle

size, temperature, pressure applied) need to be carefully controlled, and there is a need for harmonization and validation (by comparison with *in vivo* assays) of the *in vitro* digestion models used.

Regarding human bioavailability, high pressure processing increased absorption of orange juice flavanones over conventionally pasteurized orange juice [67]. No data are available for carotenoids. To our knowledge, the effect of pulsed electric fields has been investigated only on vitamin C bioavailability in humans from orange juice [68] and Mediterranean vegetable soup [69]. No studies addressing the effect of pulsed electric field on carotenoid and polyphenol bioaccessibility and bioavailability are available. Other nonthermal processing technologies such as ultrasound [70], food irradiation, light pulses or oscillating magnetic fields, have received little or no attention in terms of their effect on bioaccessibility and bioavailability of phytochemicals. Thus, more studies in this field may be required especially as more novel processing techniques are being developed.

3 Mixtures of authentic food matrices

There has been comparatively little effort into assessing the effects of characteristic combinations of foodstuffs or realistic meals on the bioaccessibility of phytochemicals. However, studies have shown that polyphenols from foods in combinations can have very different bioaccessibilities. Studies with raspberry juice showed that the addition of ice-cream markedly reduced the recovery of total anthocyanins [71], whereas a wheat-based breakfast cereal did not influence recovery. Co-digestion with blueberries \pm oat meal and/or milk revealed that milk reduced the recovery of total anthocyanins and total phenols [72] and co-digestion of nuts and dried fruits yielded lower levels of total available phenols after simulated digestion than nuts or fruits alone.

The addition of bovine, soy and rice milks, ascorbic acid or citrus juices increased the bioaccessibility of galloylated green tea catechins (EGC, EGCG, EC, and ECG), by stabilization and protection from degradation at alkaline pH [73]. The simultaneous presence of sucrose and ascorbic acid in green tea increased EGC and EGCG bioaccessibility, uptake in Caco-2 cells and bioavailability in rats [74]. Although the addition of skimmed milk reduced recovery of green tea catechins after *in vitro* digestion, the uptake of catechins by Caco-2 cells was increased [75]. This suggests that catechins bound to the milk proteins after *in vitro* digestion were available for uptake by the Caco-2 cells. This may explain the lack of difference in serum/plasma bioavailability of tea catechins in studies where subjects were given tea with or without milk [76]. Similarly, chocolate containing higher sucrose levels increased plasma concentrations of metabolites derived from catechins compared to dark and milk chocolate in rats [77]. In humans, sucrose but also solid/beverage format influenced various aspects of the bioavailability of flavan-3-ols from commercial cocoa based products [78]. Dietary fats can

increase polyphenol bioavailability in humans by increasing absorption, possibly by enhancing micellarization in the small intestine [79, 80], as noted for carotenoids. For example, higher fat content increased the stability of cocoa procyanidins during *in vitro* digestion [81]. This is particularly relevant as screening of different formulations (or different plant varieties) for the bioaccessibility of specific phytochemical components has become more common and it is possible that variations in other macronutrients influence the outcome. Using experience gained in pharmaceutical applications, strategies may also be designed to improve polyphenol bioavailability by co-administering phenolics with compounds which modulate gut and/or liver metabolizing enzymes [80, 82, 83].

There is definitely more scope for the study of food mixtures that are more realistic of real meal choices. Complex models developed by the pharmaceutical industry to assess the digestion behavior of different drug formulations [84, 85] could be applied to estimate digestion of foods. Despite the increasing complexity achieved by *in vitro* models, they remain simple compared to *in vivo* models. Typical limitations of *in vitro* models include: absence of host response factors, poor stimulation of complex mechanical forces and gastric emptying, absence of microbial flora, low level of integration into an overall digestive process, general adherence to healthy/average conditions, and limited correlation to *in vivo* situations [86]. Nevertheless, some examples have indicated a reasonable correlation between bioaccessibility studies and the situation *in vivo*. For lipophilic compounds such as α - and γ -tocopherol, β -carotene and lycopene [87], β -cryptoxanthin [88] or phytosterols [89], high correlations have been found, indicating that estimating *in vitro* bioaccessibility (solubility/micellarization) can be indicative of the amount available for uptake in the GI tract *in vivo*. For polyphenols, comparisons to *in vitro* data have been allowed by studying ileostomists [15]. In addition, combining *in vitro* digestion models and human intestinal cells (e.g. Caco-2 cells with or without a layer of mucus-producing cells such as HT-29 MTX), carotenoid uptake [90–92] qualitatively and quantitatively correlated well with human data. Investigators using *in vitro* methods must consider how to adapt the digestion conditions according to the composition of the sample and/or to food components, seeking a balance between technical simplification and accuracy, considering the *in vivo* situation as reference.

However, the release of phytochemicals from different foods is complex and involves large numbers of variables [93–95]. Furthermore, the transfer of phytochemicals from authentic food matrices will be even more complex due to a greater number of potentially rate-determining processes. Digestion and absorption rates are limited by physical processes, operating within and between the different phases of the liquid and solid phases of the digesta [96]. Using models of the human stomach [95, 97], distinct disintegration profiles and kinetics have been found for different food categories (e.g. meat products, nuts, fruits, baked and fried products,

etc.). The digestion process in terms of secretion of gastric fluids and enzymatic degradation of macronutrients is well understood, but further detailed investigations are needed to understand lipid absorption and to predict responses in realistic meals. Further information on the release of bioactive compounds should provide a better understanding of how to combine foods and design meals to improve bioavailability and enhance potential bioactive effects.

Encapsulation may enhance the bio-effectiveness of phytochemicals, and the choice of encapsulation agent can influence stability/recovery after digestion [98]. For example, Haratifar et al. [99] found that casein micelles were effective encapsulation agents for EGCG. A novel approach involving biosorption of phenolics to *Saccharomyces cerevisiae* significantly improved bioaccessibility of total phenolics, presumably as the yeast provided a protective carrier against degradation at neutral pH [100].

The possibility that certain phytochemical classes could protect other phytochemicals is an interesting, but largely under-researched area. The fact that the stability of individual phytochemical classes may be dependent on the overall phenolic composition of the sample has been raised in studies of berry polyphenol bioaccessibility [101, 102]. This interdependence could be examined in studies where blends of juices are tested for bioaccessibility of a range of phytochemical components such as vitamin C, carotenoids, and polyphenols [103]. The possibility that polyphenols may sacrificially protect carotenoids or vitamin C (or vice versa) could also be examined. For example, the effect of anthocyanins on carotenoid stability could be examined in comparative studies of purple tomatoes and related red tomatoes [104] or indeed in the recently genetically modified anthocyanin-accumulating tomatoes over their non-genetically modified counterparts [105]. Combining foodstuffs, therefore, may provide another route to enhance the bioaccessibility of specific health beneficial components. Vice versa, it has been claimed that polyphenols may modulate the digestion of macronutrients (such as starches or fats) through inhibition of their digestive enzymes [106], which could possibly modulate their own stability in the gut. The possibility that the underestimated NEPP content of fruits/vegetables [107] could influence the activity of digestive enzymes or limit the access of certain phytonutrients to digestive processes could be examined in carefully designed studies where the effect of *in vitro* digestion on the phytochemical profile of whole fruit/vegetable purees was compared to extracts and cross-compared to re-combined samples containing NEPP plus extracts.

There has been a general shift away from measuring the levels of total phytochemical classes (e.g. total carotenoids and phenolics) to targeted analysis of compositional changes in specific components during digestion, as this is more relevant to the potential health benefits attributable to specific phytochemicals. However, greater value could be obtained by using an untargeted, metabolic profiling approach which could also identify potential breakdown products and other novel components.

4 Gastric phase and small intestine

The absorption of secondary plant components includes several phases (outlined below), and prominent gaps in our knowledge include factors influencing solubilization, micelle formation (of apolar compounds), diffusion to the unstirred water layer (including the influence of mucus), transporters involved in the uptake of phytochemicals, and factors affecting phase I/II metabolism and efflux pumps.

4.1 Release from the food matrix

Aspects impeding the release of phytochemicals include large particle size [108], high meal viscosity [18] reducing the transfer of lipophilic compounds to micelles and hindering interactions between lipase and oil droplets [109]; or the presence of physically inaccessible forms such as NEPP [19,110]. As the majority of micronutrients and phytochemicals are presumably taken up in the small intestine (in their native form), and the epithelial “leakiness” decreases toward the colon [111], it is important that compounds are bioaccessible at this stage. Many polyphenols may not be detectable in the native matrix following chemical extraction, but may be released during digestion in the small intestine, such as those bound covalently or occluded by e.g. in accessible starch [112], though colonic fermentation may further result in the breakdown of NEPPs [113]. Standardized methodologies which recognize and examine the potential contribution of NEPPs are required. It also has to be noted that polyphenols have the ability to reduce the activity of digestion enzymes (e.g. pepsin, lipase), thus high concentrations of polyphenols may reduce liberation of lipids and proteins, increasing the nondigestible bulk and in turn may result in increased amounts of polyphenols passed onto the colon [19].

4.2 Solubilization and micellarization

Solubility is not an issue for most polyphenols, but more lipophilic compounds such as carotenoids [114], triterpenes [115], and phytosterols [116] require emulsification/micellarization before uptake. It remains largely unknown to what extent digestion and food matrix influence micelle formation or size. Large micelles would compromise diffusion and subsequent release of apolar compounds. Micelle diameters of approximately 6–8 nm [117,118] have been measured, but the relation of micelle size, shape, or constituents, and cellular uptake has never been studied in detail. Also, surprisingly little is known about the influence of dietary lipids and their digestion on polyphenol uptake [19], though there are indications that certain apolar polyphenols (e.g. curcumin, resveratrol, quercetin aglycones) are incorporated into micelles, and that lipid-rich foods may enhance their bioavailability [79,119–121].

A poorly understood factor in bioavailability is the role of brush border membrane enzymes, i.e. maltase, lactase-phlorizin-hydrolase, sucrase-isomaltase, and peptidases [122]. Lactase-phlorizin-hydrolase may play a crucial role in cleaving polyphenol glycosides, resulting in the uptake of free aglycones [123,124]. However, it is not certain whether cleavage occurs at this stage, by cytosolic-beta-glucosidase [19], or by colonic microbial action. The presence of esterases in the brush-border, surely present in the enterocyte [125], has been speculated on, which would influence polyphenol esters such as chlorogenic acid, or xanthophyll esters, though cholesterol esterase may also act on the latter [126].

While proteins can exert negative effects on polyphenol bioavailability [127], the interaction of apolar phytochemicals and proteins during digestion has never been studied systematically, though certain proteins may aid in the emulsification of lipid soluble phytochemicals [128]. Contrarily, a positive effect of sugars on polyphenol glucoside uptake [via stimulation of sodium-glucose linked transporter 1 (SGLT-1)] has been suggested [129]. The effect of dietary fibers is assumed to be negative, due to gel formation, enhanced viscosity, or binding and entrapping of phytochemicals, but the effects of soluble versus insoluble, or prebiotic fiber are largely unknown. Other interactions with the food matrix exist during digestion, such as high concentrations of minerals impeding micelle formation [130], but their relevance in bioavailability remains unclear.

4.3 Cellular uptake

Prior to reaching the cellular surface, diffusion through the mucus [122] is required, though properties influencing diffusion are poorly understood. Viscosity and particle/micelle size of the digesta are expected to play a role. Porcine trials have shown that smaller particles diffuse more readily through the mucus layer [131]; and it may be assumed that apolar compounds are likewise hindered [132]. More sophisticated model studies including mucus-producing cells, such as HT-29-MTX, are still rare but warranted.

Cellular uptake can occur by transcellular or paracellular routes. The latter is reserved for rather polar and small molecules <600 Da [133], i.e. ions, water, sugars, etc., as these pass through the tight junctions of the epithelium. Most phytochemicals are taken up via transcellular transport. Certain compounds (such as apple polyphenols) increase tight junction functionality [134], possibly via altered cellular signaling transduction pathways [135]. On the other hand, certain fatty acids (such as caprylic acid) and high molecular weight polyphenols reduced tight junction barrier function, and may enhance uptake of other small compounds [134,136,137].

Transcellular uptake can occur by facilitated, active means or via passive diffusion. The latter is reserved for small and apolar molecules, as these readily pass through the cell membrane [138]. Passive diffusion has been suggested for carotenoids [22,23] and some apolar polyphenol aglycones

[19]. At low concentrations, presumably similar to levels of an average mixed diet, these compounds may also be taken up by carriers, actively or by facilitated diffusion. Over 400 transmembrane carrier proteins are known [139]. Many polyphenols can inhibit these transporters, but stimulation was also described [140] and effects may depend on the polyphenol type, its concentration, exposure time, cell type, and substrate. For carotenoids, high concentrations of beta-carotene reduced lutein uptake [141], which may be explained by higher affinity to transporters such as cluster of differentiation 36 (CD36) or unknown types [142]. Enhanced uptake of polyphenol glucosides by sugars via SGLT-1 activation [19, 143] is known, but the involvement of gut transporters other than SGLT-1 and mono-carboxyl-transporter 1 (MCT1) for polyphenols is not certain, and most polyphenol aglycones are assumed to be taken up by passive diffusion [124].

4.4 First pass metabolism

A large number of metabolites can be formed following phase I (e.g. reduction/oxidation, methylation, hydroxylation, hydrolysis, e.g. via cytochrome P450-dependent mixed-function oxidases (CYPs) and catechol-O-methyltransferase) and phase II metabolism (e.g. glucuronidation by uridine-5'-diphosphate glucuronosyltransferase, and sulfation via sulfotransferases), in human enterocytes [144, 145]. Certain phytochemicals can up- or downregulate these enzymes, influencing the availability of the native compounds. For example, uptake of curcumin was approximately 20-fold enhanced when co-administered with piperine in humans [146]. High doses of certain polyphenols may override phase I/II metabolism, though not much is known on thresholds in this respect [19]. Such interactions are certainly a gap in our knowledge. For carotenoids, cleavage by beta-carotene oxygenase 1/beta-carotene dioxygenase 2 results in symmetric/asymmetric cleavage, producing apo-carotenals. However, these reactions are far from quantitative, and appear to depend on genetic factors and the type of carotenoid [23]. The bioactivities of apo-carotenals (except retinols) are poorly comprehended, though they may be highly effective, e.g. in activating nuclear receptors [147]. Detecting small amounts of these compounds is difficult, and suitable standards are often not commercially available.

4.5 Transport through the epithelium

Many phytochemicals are treated as xenobiotics, expelled from the cell, typically by increasing their polarity and via efflux transporters. These transmembrane proteins are typically ATP-dependent efflux pumps. Little is known on the specificity of these transporters, and less on the potential of phytochemicals or nutrients to block them, which could result in enhanced bioavailability. It is assumed that P-glycoprotein, MRP2 (multidrug resistance protein 2), and breast cancer re-

sistance protein (BCRP) are the most important transporters, and that these may be competitively inhibited by polyphenols, perhaps by blocking ATPase [19]. The transport from the cell to the basolateral side (i.e. bloodstream or lymph) is even more poorly understood. Compounds that could increase or decrease MRP3 (or MRP1, MRP4, and possibly several MCTs), could affect polyphenol uptake, as e.g. demonstrated by mice overexpressing MRP3 and showing high resveratrol bioavailability [148]. It is assumed that carotenoids can be transported back into the lumen by scavenger receptor class B member 1 and possibly ATP-binding cassette subfamily G, member 5/8, but more research is needed. Transport through the cells, e.g. by transporters such as fatty acid binding proteins is assumed, but not confirmed [23]. It can be further assumed that proteins involved in chylomicron generation [i.e. microsomal triglyceride transfer protein, apoB48, apoAIV, and Sar1b [23, 149], also influence carotenoid transport. For polyphenols, transport through the cells is not understood, and may occur primarily by diffusion.

5 Metabolism in the colon and other organs

5.1 Metabolism of polyphenols

In general, polyphenol metabolism is fairly well known due to their xenobiotic nature. Polyphenols undergo metabolism in intestinal and liver tissues and by colon microbiota [15]. In fact, it seems that the colon may be the major important site for polyphenol uptake, at least for orange juice rich in hesperitin and naringin [150], and studies with ileostomists suggested significant absorption from the colon [151]. Phenolic compounds are glucuronidated and sulfated in the liver and intestinal tissues, and these metabolites are found in body fluids [125, 152–155]. Hepatic metabolites can be recycled back to the small intestine through biliary excretion [27, 156–159] and end up in the colon, where they are deglucuronidated by microbial α , D-glucuronidases before ring fission [160, 161].

Sophisticated methods for studying enterohepatic circulation, including humans, using a perfusion technique, have been published [162, 163], in addition to various articles on tissue metabolites of phenolic compounds [164], e.g. flavanol monomers and tea polyphenols. However, the enterohepatic circulation of colonic metabolites requires further investigation and hepatic metabolism of colonic metabolites of plant phenolic compounds should be addressed in the future [162]. Furthermore, Monagas et al. [165] suggested that microbial metabolites may act as signal molecules, and their action should be taken into account in more detail in future investigations.

Absorbed microbial-derived metabolites (such as enterolignans and dihydroxylated compounds such as methyl catechol) can be subjected to further glucuronidation, methylation, sulfation, or glycation in the liver, while phase I metabolism (oxidation/reduction reactions) seems to occur

to a lesser extent [27]. The interplay between the liver and the colon, the enterohepatic circulation, leads to a long residence time (up to 24–48 h) in the blood [157, 162, 166–168], which can go unobserved unless longer sampling of blood or urine is carried out [164, 169]. Finally, the phenolic microbial metabolites are distributed to tissues and are excreted via urine, partly as free but mainly as hepatic conjugates, depending on the structure of the parent backbone [125, 157, 158].

One of the major gaps is the lack of knowledge concerning hepatic conversion of small colon-derived phenolic acids, which may be missed in analyses of human body fluids or cell lines due to their hydrophilic nature. Extraction of samples may also cause bias, as Sawai et al. [157] elucidated the ratio of excreted conjugated and free phenolic acid colon-derived metabolites from quercetin derivatives in urine, by using water saturated ethyl acetate to enhance the yield of polar conjugates.

Although human studies are more relevant, mechanisms of action can be studied using *in vitro* human-based cell model systems designed to study properties of drugs [27]. In recent years, novel cell lines and culture strategies have helped in overcoming the scarcity of human liver material and problems in maintaining the expression and function of metabolizing enzymes [170]. The advent of human stem cell derived hepatocytes will potentially provide an unlimited source of human hepatocytes [171]. Human hepatocyte three-dimensional models, with complete hepatic metabolizing enzymes, transporters, and cofactors, may be applicable to metabolite profiling, pathway identification, CYP450 inhibition, CYP450 induction, and uptake and efflux transporter inhibition by polyphenols and their metabolites.

The lack of standards for metabolites causes limitations to study both polyphenol and phytosterol metabolism. Chemical procedures and biochemical labeling tools are available for the synthesis of many conjugated metabolites [172]. However, the absence of standards limits the use of some analytical methodologies, and frequently enzyme treatment is used to overcome the problem of hepatic conjugative metabolism. For example, conjugated flavan-3-ol metabolites are rarely available commercially, and most studies have analyzed plasma and urine samples after treatment with glucuronidase/sulfatase, providing data only for the aglycones [164]. However, some sulfate conjugates are resistant to enzyme hydrolysis, thus, this methodology may underestimate bioavailability [173, 174]. This is the situation for estimating epicatechin bioavailability from cocoa products. Direct analysis of the individual epicatechin metabolites by LC-MS/MS may overcome this problem, however, the chirality of the aglycone will still be unknown, and it is plausible that the enantiomers could differ in their biological activity. Each flavan-3-ol and each associated phase II conjugate can, in theory, occur as four enantiomers, with the (+) and (–) forms resolvable only by chiral chromatography, and so far as we are aware there are no reports of chiral analysis of conjugated metabolites in urine or plasma [164].

However, targeted LC-MS analytical approaches can be used for identifying many metabolites by exact mass, if a theoretical prediction of possible metabolites and conjugates based on expected metabolism is available [154]. Nontargeted metabolomic profiling can find metabolites based on structural similarity to the parent compound, if the mass spectra can be coupled with a compound library. Profiling of phenolic metabolomes has been assessed by NMR, GC, LC and 2D-GC with TOF mass detection coupled with a compound library [25, 175–178].

5.2 Metabolism of lipophilic compounds

Regarding lipophilic bioactive compounds, serum phytosterol bioavailability is reported to be below 10%, suggesting that they may reach the colon and subjected to microbial metabolism [179]. The pathway of microbial transformation from sterol to stanol form in humans was reported in the 1970's, but the kinetics of the reaction steps and metabolites in the large intestine have not been studied systematically [180]. For carotenoids, the liver stores and distributes carotenoids to other tissues, but the mechanisms are not known accurately [181]. However, intervention studies have shown that increased dietary intake of carotenoids influences serum concentrations more than colon concentrations [182], suggesting that absorption occurs mostly in the small intestine. In addition, metabolites of lycopene produced *in vivo* also occur naturally at low concentrations in tomato, which causes difficulties in the differentiation of the origin of the metabolites [183].

5.3 Transport, tissue distribution, and mechanism of action

Transport of metabolites and their parent compounds to tissues has not been studied adequately. This is important, since it is the key action to ensure the desired biological activity. There is a lack of information whether conjugated polyphenols can prolong their residence time in the body by modulating the activity of the transporters in the excretory organs. A recent study of transmembrane transport of flavonoids and some of their methylated and glucuronidated metabolites using human cerebral microvessel endothelial cells as a blood–brain barrier cell model [184] showed that the metabolites were transported in a time-dependent manner and showed higher transport efficiency than the native flavonoids and were not further transformed by the cells. Polyphenols affect their own metabolism and tissue distribution, due to their capacity to modulate the activity of xenobiotic metabolizing enzymes and transporters, modulating their own concentration-time profiles in the body and also promoting alterations in drug or toxin pharmacokinetics, the mechanisms of which are not known [140, 185]. Therefore, additive, synergistic, or antagonistic effects of xenobiotics (dietary

phytochemicals, drugs, and toxins) need to be addressed. For example, the downregulation of cytochrome P450 3A4 by grapefruit juice and its polyphenols, enhancing the concentration of several pharmaceuticals and increasing their effects is well known [186]. Furthermore, the knowledge of pharmacodynamics and tissue distribution for most polyphenols and their metabolites is still lacking. It would also be relevant to study individual variation in the tissue distribution and the potential bioactivity caused by genetic variation in pathways related to bioavailability and transport [187].

In vitro models for preclinical research using stem cells, and patient-specific induced pluripotent stem cells and reprogrammed somatic cells from patients are already applied in disease modeling and drug discovery, and may be applicable to test polyphenol metabolite health benefits. Micro-engineered physiological systems, also known as “organs-on-chips”, can reconstitute physiologically critical features of human tissues and their interactions [188]. The nematode *Caenorhabditis elegans*, with the presence of tissue and organ systems, is increasingly used as an in vivo model and has been employed to study the metabolism of methylated catechin derivatives and their biological effects on oxidative and thermal stress resistance [189]. Thus, novel cell assays and nematode in vivo assays may be applied to study mechanisms of action of the parent compounds versus their liver/colon metabolites, although it should be noted that in vivo models such as *C. elegans* or *Drosophila* (like mice) may produce quite different metabolites from ingested phytochemicals than humans.

Only a few studies have combined the assay of biological effects of bioactive compounds or their colonic metabolites from whole foods or their extracts with an in vitro GI digestion process with/without colonic fermentation [102, 172, 190], or compared the biological activity of metabolites to their parent compound [178]. Furthermore, the synergistic effects of the phytochemical metabolite pool, including the interplay between liver and colon microbiota should be studied. Finally, cell-based assays measuring the bioactivity of the metabolites at relevant concentration are scarce [102, 191, 192]. In the future, bioactivity assays to mimic the actual in vivo situation in the corresponding tissue, should use metabolite pools from whole foods, at tissue-tolerant concentrations.

6 Conclusion

In this position paper, we have tried to highlight gaps of knowledge with respect to selected phytochemicals. It has to be noted that the data presented represent a somewhat simple and general paradigm for phytochemicals, and that, due to the very large variety and properties of secondary plant compounds, the statements cannot be generalized toward all groups of phytochemicals. In addition, some of the missing aspects discussed in this paper may currently be tackled, but have not been published.

However, while recent work has greatly enhanced our insight into the metabolism and bioavailability of a range of phytochemicals, many factors governing matrix release, solubilization, cellular uptake, and biotransformation remain poorly understood. Major aspects which deserve more attention when estimating bioavailability aspects include effects of innovative processing techniques, synergistic effects of mixed/whole diets, factors effecting micelle formation, co-constituents influencing influx and efflux via transporter systems or altering phase I/II metabolism, as these have often been overlooked or excluded from consideration, in part due to difficulties to include their study in vivo or in vitro. In the future, enhanced availability of analytical possibilities to investigate these aspects such as through broader availability of instruments to measure food texture, visualization of micelles (TEM, Mastersizer), improved cell models of absorption and metabolism (mucus producing, liver cells, 3D models), ways to produce knock-out variants (e.g. of certain transporters) in animal models (nematodes, mice, etc.) to study pathways of absorption and bioactivity of metabolites, and improved chromatographic techniques and commercial availability of metabolites (e.g. sulfates and glucuronides) will aid toward an improved understanding of these important aspects of bioavailability.

The EU-COST FA 1005 Infogest action is greatly acknowledged for providing funding for travel and meetings and allowing for open access.

The authors have declared no conflict of interest.

7 References

- [1] Hooper, L., Kroon, P. A., Rimm, E. B., Cohn, J. S. et al., Flavonoids, flavonoid-rich foods, and cardiovascular risk: a meta-analysis of randomized controlled trials. *Am. J. Clin. Nutr.* 2008, *88*, 38–50.
- [2] Hamer, M., Chida, Y., Intake of fruit, vegetables, and antioxidants and risk of type 2 diabetes: systematic review and meta-analysis. *J. Hypertens.* 2007, *25*, 2361–2369.
- [3] Bar-Sela, G., Epelbaum, R., Schaffer, M., Curcumin as an anti-cancer agent: review of the gap between basic and clinical applications. *Curr. Med. Chem.* 2010, *17*, 190–197.
- [4] Hausenblas, H. A., Schoulda, J. A., Smoliga, J. M., Resveratrol treatment as an adjunct to pharmacological management in type 2 diabetes mellitus—systematic review and meta-analysis. *Mol. Nutr. Food Res.* 2015, *59*, 147–159.
- [5] Jacobs, D. R., Tapsell, L. C., Food synergy: the key to a healthy diet. *Proc. Nutr. Soc.* 2013, *72*, 200–206.
- [6] Prasain, J. K., Carlson, S. H., Wyss, J. M., Flavonoids and age-related disease: risk, benefits and critical windows. *Maturitas* 2010, *66*, 163–171.
- [7] Bjelakovic, G., Nikolova, D., Gluud, L. L., Simonetti, R. G. et al., Mortality in randomized trials of antioxidant supplements for primary and secondary prevention: systematic review and meta-analysis. *JAMA* 2007, *297*, 842–857.

- [8] Bjelakovic, G., Nikolova, D., Gluud, C., Antioxidant supplements and mortality. *Curr. Opin. Clin. Nutr. Metab. Care* 2014, *17*, 40–44.
- [9] Ye, Y., Li, J., Yuan, Z., Effect of antioxidant vitamin supplementation on cardiovascular outcomes: a meta-analysis of randomized controlled trials. *PLoS One* 2013, *8*, e56803.
- [10] Jordan, V. C., Avoiding the bad and enhancing the good of soy supplements in breast cancer. *J. Natl. Cancer Inst.* 2014, *106*, 1–3.
- [11] Del, R. D., Rodriguez-Mateos, A., Spencer, J. P., Tognolini, M. et al., Dietary (poly)phenolics in human health: structures, bioavailability, and evidence of protective effects against chronic diseases. *Antioxid. Redox Signal.* 2013, *18*, 1818–1892.
- [12] Moran, N. E., Erdman, J. W., Jr., Clinton, S. K., Complex interactions between dietary and genetic factors impact lycopene metabolism and distribution. *Arch. Biochem. Biophys.* 2013, *539*, 171–180.
- [13] Rodriguez-Mateos, A., Vauzour, D., Krueger, C. G., Shanmuganayagam, D. et al., Bioavailability, bioactivity and impact on health of dietary flavonoids and related compounds: an update. *Arch. Toxicol.* 2014, *88*, 1803–1853.
- [14] Holst, B., Williamson, G., Nutrients and phytochemicals: from bioavailability to bioefficacy beyond antioxidants. *Curr. Opin. Biotechnol.* 2008, *19*, 73–82.
- [15] Alminger, M., Aura, A.-M., Bohn, T., Dufour, C. et al., In vitro models for studying secondary plant metabolite digestion and bioaccessibility. *Compr. Rev. Fd. Technol. Fd. Saf.* 2014, *13*, 413–436.
- [16] Svelander, C. A., Lopez-Sanchez, P., Pudney, P. D., Schumm, S. et al., High pressure homogenization increases the in vitro bioaccessibility of alpha- and beta-carotene in carrot emulsions but not of lycopene in tomato emulsions. *J. Food Sci.* 2011, *76*, H215–H225.
- [17] Wang, L., Bohn, T., in: Bouayed, J., Bohn, T. (Eds.), *Nutrition, Well-Being and Health*, Intech, Croatia 2012, pp. 201–224.
- [18] Panozzo, A., Lemmens, L., Van, L. A., Manzocco, L. et al., Microstructure and bioaccessibility of different carotenoid species as affected by high pressure homogenisation: a case study on differently coloured tomatoes. *Food Chem.* 2013, *141*, 4094–4100.
- [19] Bohn, T., Dietary factors affecting polyphenol bioavailability. *Nut. Rev.* 2014, *72*, 429–452.
- [20] Minekus, M., Alminger, M., Alvito, P., Ballance, S. et al., A standardised static in-vitro digestion method suitable for food – an international consensus. *Food Funct.* 2014, *5*, 1113–1124.
- [21] Czank, C., Cassidy, A., Zhang, Q., Morrison, D. J. et al., Human metabolism and elimination of the anthocyanin, cyanidin-3-glucoside: a (13)C-tracer study. *Am. J. Clin. Nutr.* 2013, *97*, 995–1003.
- [22] Bohn, T., Bioavailability of non-provitamin A carotenoids. *Curr. Nutr. Food Sci.* 2008, *4*, 240–258.
- [23] Borel, P., Genetic variations involved in interindividual variability in carotenoid status. *Mol. Nutr. Food Res.* 2012, *56*, 228–240.
- [24] Bazzocco, S., Mattila, I., Guyot, S., Renard, C. M. et al., Factors affecting the conversion of apple polyphenols to phenolic acids and fruit matrix to short-chain fatty acids by human faecal microbiota in vitro. *Eur. J. Nutr.* 2008, *47*, 442–452.
- [25] Aura, A. M., Mattila, I., Hyotylainen, T., Gopalacharyulu, P. et al., Characterization of microbial metabolism of Syrah grape products in an in vitro colon model using targeted and non-targeted analytical approaches. *Eur. J. Nutr.* 2013, *52*, 833–846.
- [26] Aura, A. M., Mattila, I., Seppainen-Laakso, T., Miettinen, J. et al., Microbial metabolism of catechin stereoisomers by human faecal microbiota: comparison of targeted analysis and a non-targeted metabolomics method. *Phytochem. Lett.* 2008, *1*, 18–22.
- [27] del Rio, D., Rodriguez-Mateos, A., Spencer, J. P., Tognolini, M. et al., Dietary (poly)phenolics in human health: structures, bioavailability, and evidence of protective effects against chronic diseases. *Antioxid. Redox. Signal.* 2013, *18*, 1818–1892.
- [28] Verkerk, R., Schreiner, M., Krumbein, A., Ciska, E. et al., Glucosinolates in Brassica vegetables: the influence of the food supply chain on intake, bioavailability and human health. *Mol. Nutr. Food Res.* 2009, *53*(Suppl 2), S219–S265.
- [29] Van Buggenhout, S., Alminger, M., Lemmens, L., Colle, I. et al., In vitro approaches to estimate the effect of food processing on carotenoid bioavailability need thorough understanding of process induced microstructural changes. *Trends Food Sci. Technol.* 2010, *21*, 607–618.
- [30] Sensoy, I., A review on the relationship between food structure, processing, and bioavailability. *Crit. Rev. Food Sci. Nutr.* 2014, *54*, 902–909.
- [31] Schweiggert, R. M., Mezger, D., Schimpf, F., Steingass, C. B. et al., Influence of chromoplast morphology on carotenoid bioaccessibility of carrot, mango, papaya, and tomato. *Food Chem.* 2012, *135*, 2736–2742.
- [32] Tydeman, E. A., Parker, M. L., Faulks, R. M., Cross, K. L. et al., Effect of carrot (*Daucus carota*) microstructure on carotene bioaccessibility in the upper gastrointestinal tract. 2. In vivo digestions. *J. Agric. Food Chem.* 2010, *58*, 9855–9860.
- [33] Palafox-Carlos, H., yala-Zavala, J. F., Gonzalez-Aguilar, G. A., The role of dietary fiber in the bioaccessibility and bioavailability of fruit and vegetable antioxidants. *J. Food Sci.* 2011, *76*, R6–R15.
- [34] Buchweitz, M., Speth, M., Kammerer, D. R., Carle, R., Stabilisation of strawberry (*Fragaria x ananassa* Duch.) anthocyanins by different pectins. *Food Chem.* 2013, *141*, 2998–3006.
- [35] Kodera, Y., Suzuki, A., Imada, O., Kasuga, S. et al., Physical, chemical, and biological properties of s-allylcysteine, an amino acid derived from garlic. *J. Agric. Food Chem.* 2002, *50*, 622–632.
- [36] Tapiero, H., Townsend, D. M., Tew, K. D., Organosulfur compounds from alliaceae in the prevention of human pathologies. *Biomed. Pharmacother.* 2004, *58*, 183–193.

- [37] Holst, B., Williamson, G., A critical review of the bioavailability of glucosinolates and related compounds. *Nat. Prod. Rep.* 2004, *21*, 425–447.
- [38] Ioannou, I., Hafsa, I., Hamdi, S., Charbonnel, C. Å. et al., Review of the effects of food processing and formulation on flavonol and anthocyanin behaviour. *J. Food Engin.* 2012, *111*, 208–217.
- [39] Reif, C., Arrigoni, E., Berger, F., Baumgartner, D. et al., Lutein and beta-carotene content of green leafy Brassica species grown under different conditions. *LWT - Food Sci. Technol.* 2013, *53*, 378–381.
- [40] Szydłowska-Czerniak, A., Trokowski, K., Karlovits, G., Szlyk, E., Effect of refining processes on antioxidant capacity, total contents of phenolics and carotenoids in palm oils. *Food Chem.* 2011, *129*, 1187–1192.
- [41] Szydłowska-Czerniak, A., Karlovits, G., Dianoczki, C., Recseg, K. et al., Comparison of two analytical methods for assessing antioxidant capacity of rapeseed and olive oils. *J. Am. Oil Chem. Soc.* 2008, *85*, 141–149.
- [42] Wang, C., Riedl, K. M., Schwartz, S. J., Fate of folates during vegetable juice processing – deglutamylation and interconversion. *Food Res. Int.* 2013, *53*, 440–448.
- [43] Hung, P. V., Maeda, T., Miyatake, K., Morita, N., Total phenolic compounds and antioxidant capacity of wheat graded flours by polishing method. *Food Res. Int.* 2009, *42*, 185–190.
- [44] Pradeep, S. R., Guha, M., Effect of processing methods on the nutraceutical and antioxidant properties of little millet (*Panicum sumatrense*) extracts. *Food Chem.* 2011, *126*, 1643–1647.
- [45] Kim, J. S., Kang, O. J., Gweon, O. C., Comparison of phenolic acids and flavonoids in black garlic at different thermal processing steps. *J. Funct. Foods* 2013, *5*, 80–86.
- [46] Wu, L., Huang, Z., Qin, P., Ren, G., Effects of processing on phytochemical profiles and biological activities for production of sorghum tea. *Food Res. Int.* 2013, *53*, 678–685.
- [47] Leong, S. Y., Oey, I., Effects of processing on anthocyanins, carotenoids and vitamin C in summer fruits and vegetables. *Food Chem.* 2012, *133*, 1577–1587.
- [48] Karakaya, S., Yilmaz, N., Lycopene content and antioxidant activity of fresh and processed tomatoes and in vitro bioavailability of lycopene. *J. Sci. Food Agric.* 2007, *87*, 2342–2347.
- [49] Sommano, S., Caffin, N., McDonald, J., Cocksedge, R., The impact of thermal processing on bioactive compounds in Australian native food products (bush tomato and Kakadu plum). *Food Res. Int.* 2013, *50*, 557–561.
- [50] Zhang, D., Hamazu, Y., Phenolics, ascorbic acid, carotenoids and antioxidant activity of broccoli and their changes during conventional and microwave cooking. *Food Chem.* 2004, *88*, 503–509.
- [51] Mayer-Miebach, E., Behnsilian, D., Regier, M., Schuchmann, H. P., Thermal processing of carrots: lycopene stability and isomerisation with regard to antioxidant potential. *Food Res. Int.* 2005, *38*, 1103–1108.
- [52] Mayeaux, M., Xu, Z., King, J. M., Prinyawiwatkul, W., Effects of cooking conditions on the lycopene content in tomatoes. *J. Food Sci.* 2006, *71*, C461–C464.
- [53] Seybold, C., Frohlich, K., Bitsch, R., Otto, K. et al., Changes in contents of carotenoids and vitamin E during tomato processing. *J. Agric. Food Chem.* 2004, *52*, 7005–7010.
- [54] Pugliese, A., Loizzo, M. R., Tundis, R., O’Callaghan, Y. et al., The effect of domestic processing on the content and bioaccessibility of carotenoids from chili peppers (*Capsicum* species). *Food Chem.* 2013, *141*, 2606–2613.
- [55] Ciska, E., Kozłowska, H., The effect of cooking on the glucosinolates content in white cabbage. *Eur. Food Res. Technol.* 2001, *212*, 582–587.
- [56] Wennberg, M., Ekvall, J., Olsson, K., Nyman, M., Changes in carbohydrate and glucosinolate composition in white cabbage (*Brassica oleracea* var. *capitata*) during blanching and treatment with acetic acid. *Food Chem.* 2006, *95*, 226–236.
- [57] Volden, J., Borge, G. I., Bengtsson, G. B., Hansen, M. et al., Effect of thermal treatment on glucosinolates and antioxidant-related parameters in red cabbage (*Brassica oleracea* L. ssp. *capitata* f. *rubra*). *Food Chem.* 2008, *109*, 595–605.
- [58] Francisco, M., Velasco, P., Moreno, D. A., Garcia-Viguera, C. et al., Cooking methods of *Brassica rapa* affect the preservation of glucosinolates, phenolics and vitamin C. *Food Res. Int.* 2010, *43*, 1455–1463.
- [59] Hanschen, F. S., Rohn, S., Mewis, I., Schreiner, M. et al., Influence of the chemical structure on the thermal degradation of the glucosinolates in broccoli sprouts. *Food Chem.* 2012, *130*, 1–8.
- [60] Girgin, N., El, S. N., Effects of cooking on in vitro sinigrin bioaccessibility, total phenols, antioxidant and antimutagenic activity of cauliflower (*Brassica oleracea* L. var. *Botrytis*). *J. Food Compos. Anal.* 2015, *37*, 119–127.
- [61] Sanchez-Moreno, C., de, A. B., Plaza, L., Elez-Martinez, P. et al., Nutritional approaches and health-related properties of plant foods processed by high pressure and pulsed electric fields. *Crit. Rev. Food Sci. Nutr.* 2009, *49*, 552–576.
- [62] McInerney, J. K., Seccafien, C. A., Stewart, C. M., Bird, A. R., Effects of high pressure processing on antioxidant activity, and total carotenoid content and availability, in vegetables. *Innov. Food Sci. Emerg. Technol.* 2007, *8*, 543–548.
- [63] Colle, I., Van Buggenhout, S., Van Loey, A., Hendrickx, M., High pressure homogenization followed by thermal processing of tomato pulp: influence on microstructure and lycopene in vitro bioaccessibility. *Food Res. Int.* 2010, *43*, 2193–2200.
- [64] Knockaert, G., Pulissery, S. K., Colle, I., Van, B. S. et al., Lycopene degradation, isomerization and in vitro bioaccessibility in high pressure homogenized tomato puree containing oil: effect of additional thermal and high pressure processing. *Food Chem.* 2012, *135*, 1290–1297.
- [65] Cilla, A., Alegria, A., de, A. B., Sanchez-Moreno, C. et al., Bioaccessibility of tocopherols, carotenoids, and ascorbic acid from milk- and soy-based fruit beverages: influence of food matrix and processing. *J. Agric. Food Chem.* 2012, *60*, 7282–7290.

- [66] Gupta, R., Kopec, R. E., Schwartz, S. J., Balasubramanian, V. M., Combined pressure-temperature effects on carotenoid retention and bioaccessibility in tomato juice. *J. Agric. Food Chem.* 2011, *59*, 7808–7817.
- [67] Tomas-Navarro, M., Vallejo, F., Sentandreu, E., Navarro, J. L. et al., Volunteer stratification is more relevant than technological treatment in orange juice flavanone bioavailability. *J. Agric. Food Chem.* 2013, *62*, 24–27.
- [68] Sanchez-Moreno, C., Cano, M. P., de, A. B., Plaza, L. et al., Pulsed electric fields-processed orange juice consumption increases plasma vitamin C and decreases F2-isoprostanes in healthy humans. *J. Nutr. Biochem.* 2004, *15*, 601–607.
- [69] Sanchez-Moreno, C., Pilar, C. M., de, A. B., Plaza, L. et al., Intake of Mediterranean vegetable soup treated by pulsed electric fields affects plasma vitamin C and antioxidant biomarkers in humans. *Int. J. Food Sci. Nutr.* 2005, *56*, 115–124.
- [70] Anese, M., Mirolo, G., Beraldo, P., Lippe, G., Effect of ultrasound treatments of tomato pulp on microstructure and lycopene in vitro bioaccessibility. *Food Chem.* 2013, *136*, 458–463.
- [71] McDougall, G. J., Dobson, P., Smith, P., Blake, A. et al., Assessing potential bioavailability of raspberry anthocyanins using an in vitro digestion system. *J. Agric. Food Chem.* 2005, *53*, 5896–5904.
- [72] Cebeci, F., Sahin-Yesilcubuk, N., The matrix effect of blueberry, oat meal and milk on polyphenols, antioxidant activity and potential bioavailability. *Int. J. Food Sci. Nutr.* 2014, *65*, 69–78.
- [73] Green, R. J., Murphy, A. S., Schulz, B., Watkins, B. A. et al., Common tea formulations modulate in vitro digestive recovery of green tea catechins. *Mol. Nutr. Food Res.* 2007, *51*, 1152–1162.
- [74] Peters, C. M., Green, R. J., Janle, E. M., Ferruzzi, M. G., Formulation with ascorbic acid and sucrose modulates catechin bioavailability from green tea. *Food Res. Int.* 2010, *43*, 95–102.
- [75] Xie, Y., Kosinska, A., Xu, H., Andlauer, W., Milk enhances intestinal absorption of green tea catechins in in vitro digestion/Caco-2 cells model. *Food Res. Int.* 2013, *53*, 793–800.
- [76] van het Hof, K. H., Kivits, G. A., Weststrate, J. A., Tijburg, L. B., Bioavailability of catechins from tea: the effect of milk. *Eur. J. Clin. Nutr.* 1998, *52*, 356–359.
- [77] Neilson, A. P., Sapper, T. N., Janle, E. M., Rudolph, R. et al., Chocolate matrix factors modulate the pharmacokinetic behavior of cocoa flavan-3-ol phase II metabolites following oral consumption by Sprague-Dawley rats. *J. Agric. Food Chem.* 2010, *58*, 6685–6691.
- [78] Neilson, A. P., George, J. C., Janle, E. M., Mattes, R. D. et al., Influence of chocolate matrix composition on cocoa flavan-3-ol bioaccessibility in vitro and bioavailability in humans. *J. Agric. Food Chem.* 2009, *57*, 9418–9426.
- [79] Guo, Y., Mah, E., Davis, C. G., Jalili, T. et al., Dietary fat increases quercetin bioavailability in overweight adults. *Mol. Nutr. Food Res.* 2013, *57*, 896–905.
- [80] Rein, M. J., Renouf, M., Cruz-Hernandez, C., Actis-Goretta, L. et al., Bioavailability of bioactive food compounds: a challenging journey to bioefficacy. *Br. J. Clin. Pharmacol.* 2013, *75*, 588–602.
- [81] Ortega, N., Reguant, J., Romero, M. P., Macia, A. et al., Effect of fat content on the digestibility and bioaccessibility of cocoa polyphenol by an in vitro digestion model. *J. Agric. Food Chem.* 2009, *57*, 5743–5749.
- [82] Henning, S. M., Choo, J. J., Heber, D., Nongallated compared with gallated flavan-3-ols in green and black tea are more bioavailable. *J. Nutr.* 2008, *138*, 1529S–1534S.
- [83] Scheepens, A., Tan, K., Paxton, J. W., Improving the oral bioavailability of beneficial polyphenols through designed synergies. *Genes Nutr.* 2010, *5*, 75–87.
- [84] Huang, W., Lee, S. L., Yu, L. X., Mechanistic approaches to predicting oral drug absorption. *AAPS J.* 2009, *11*, 217–224.
- [85] Parrott, N., Lave, T., Applications of physiologically based absorption models in drug discovery and development. *Mol. Pharm.* 2008, *5*, 760–775.
- [86] Guerra, A., Etienne-Mesmin, L., Livrelli, V., Denis, S. et al., Relevance and challenges in modeling human gastric and small intestinal digestion. *Trends Biotechnol.* 2012, *30*, 591–600.
- [87] Reboul, E., Richelle, M., Perrot, E., smoulin-Malezet, C. et al., Bioaccessibility of carotenoids and vitamin E from their main dietary sources. *J. Agric. Food Chem.* 2006, *54*, 8749–8755.
- [88] Granado-Lorencio, F., Donoso-Navarro, E., Sanchez-Siles, L. M., Blanco-Navarro, I. et al., Bioavailability of beta-cryptoxanthin in the presence of phytosterols: in vitro and in vivo studies. *J. Agric. Food Chem.* 2011, *59*, 11819–11824.
- [89] Garcia-Llatas, G., Cilla, A., Alegria, A., Lagarda, M. A. J., Bioavailability of plant sterol-enriched milk-based fruit beverages: in vivo and in vitro studies. *J. Funct. Foods* 2015, *14*, 44–50.
- [90] Garrett, D. A., Failla, M. L., Sarama, R. J., Development of an in vitro digestion method to assess carotenoid bioavailability from meals. *J. Agric. Food Chem.* 1999, *47*, 4301–4309.
- [91] Garrett, D. A., Failla, M. L., Sarama, R. J., Estimation of carotenoid bioavailability from fresh stir-fried vegetables using an in vitro digestion/Caco-2 cell culture model. *J. Nutr. Biochem.* 2000, *11*, 574–580.
- [92] Alminger, M., Svelander, C., Wellner, A., Martinez-Tomas, R. et al., Applicability of in vitro models in predicting the in vivo bioavailability of lycopene and beta-carotene from differently processed soups. *Food Nutr. Sci.* 2012, *3*, 477–489.
- [93] Van Buggenhout, S., Sila, D. N., Duvetter, T., Van Loey, A. et al., Pectins in processed fruits and vegetables: Part III “Texture Engineering” *Comp. Rev. Food Sci. Food Saf.* 2009, *8*, 105–117.
- [94] Kong, F., Singh, R. P., A model stomach system to investigate disintegration kinetics of solid foods during gastric digestion. *J. Food Sci.* 2008, *73*, E202–E210.

- [95] Kong, F., Singh, R. P., Modes of disintegration of solid foods in simulated gastric environment. *Food Biophys.* 2009, *4*, 180–190.
- [96] Lentle, R. G., Janssen, P. W. M., *The Physical Processes of Digestion*; Springer, New York 2011.
- [97] Kong, F., Oztop, M. H., Paul Singh, R., McCarthy, M. J., Effect of boiling, roasting and frying on disintegration of peanuts in simulated gastric environment. *LWT - Food Sci. Technol.* 2013, *50*, 32–38.
- [98] Flores, F. P., Singh, R. K., Kerr, W. L., Pegg, R. B. et al., Total phenolics content and antioxidant capacities of microencapsulated blueberry anthocyanins during in vitro digestion. *Food Chem.* 2014, *153*, 272–278.
- [99] Haratifar, S., Meckling, K. A., Corredig, M., Bioefficacy of tea catechins encapsulated in casein micelles tested on a normal mouse cell line (4D/WT) and its cancerous counterpart (D/v-src) before and after in vitro digestion. *Food Funct.* 2014, *5*, 1160–1166.
- [100] Jilani, H., Cilla A., Barbera R., Hamdi, M., Impact of biosorption into *Saccharomyces cerevisiae* and in vitro gastrointestinal digestion on total polyphenols and antioxidant capacity of tea and olive leaves infusions. Proceedings of the 2nd International Conference on Food Digestion, 137. *Proceedings of the 2nd International Conference on Food Digestion*, 137. 2013.
- [101] McDougall, G., Dobson, P., Shapiro, F., Smith, P. et al., Assessing bioavailability of soft fruit polyphenols in vitro. *Acta Hort.* 2007, *744*, 135–148.
- [102] Brown, E. M., McDougall, G. J., Stewart, D., Pereira-Caro, G. et al., Persistence of anticancer activity in berry extracts after simulated gastrointestinal digestion and colonic fermentation. *PLoS One* 2012, *7*, e49740.
- [103] Rodriguez-Roque, M. J., Rojas-Grau, M. A., Elez-Martinez, P., Martin-Belloso, O., Changes in vitamin C, phenolic, and carotenoid profiles throughout in vitro gastrointestinal digestion of a blended fruit juice. *J. Agric. Food Chem.* 2013, *61*, 1859–1867.
- [104] Li, H., Deng, Z., Liu, R., Loewen, S. et al., Bioaccessibility, in vitro antioxidant activities and in vivo anti-inflammatory activities of a purple tomato (*Solanum lycopersicum* L.). *Food Chem.* 2014, *159*, 353–360.
- [105] Bassolino, L., Zhang, Y., Schoonbeek, H. J., Kiferle, C. et al., Accumulation of anthocyanins in tomato skin extends shelf life. *New Phytol.* 2013, *200*, 650–655.
- [106] McDougall, G. J., Kulkarni, N. N., Stewart, D., Current developments on the inhibitory effects of berry polyphenols on digestive enzymes. *Biofactors* 2008, *34*, 73–80.
- [107] Arranz, S., Silvan, J. M., Saura-Calixto, F., Nonextractable polyphenols, usually ignored, are the major part of dietary polyphenols: a study on the Spanish diet. *Mol. Nutr. Food Res.* 2010, *54*, 1646–1658.
- [108] Maeda-Yamamoto, M., Ema, K., Tokuda, Y., Monobe, M. et al., Effect of green tea powder (*Camellia sinensis* L. cv. Benifuuki) particle size on O-methylated EGCG absorption in rats; The Kakegawa Study. *Cytotechnology* 2011, *63*, 171–179.
- [109] McClements, D. J., Decker, E. A., Park, Y., Controlling lipid bioavailability through physicochemical and structural approaches. *Crit. Rev. Food Sci. Nutr.* 2009, *49*, 48–67.
- [110] Bravo, L., Polyphenols: chemistry, dietary sources, metabolism, and nutritional significance. *Nutr. Rev.* 1998, *56*, 317–333.
- [111] Anderson, J. M., Van Itallie, C. M., Physiology and function of the tight junction. *Cold Spring Harb. Perspect. Biol.* 2009, *1*, a002584.
- [112] Miranda, L., Deusser, H., Evers, D., The impact of in vitro digestion on bioaccessibility of polyphenols from potatoes and sweet potatoes and their influence on iron absorption by human intestinal cells. *Food Funct.* 2013, *4*, 1595–1601.
- [113] Perez-Jimenez, J., AZ-Rubio, M. E., Saura-Calixto, F., Non-extractable polyphenols, a major dietary antioxidant: occurrence, metabolic fate and health effects. *Nutr. Res. Rev.* 2013, *26*, 118–129.
- [114] Chitchumroonchokchai, C., Failla, M. L., Hydrolysis of zeaxanthin esters by carboxyl ester lipase during digestion facilitates micellarization and uptake of the xanthophyll by Caco-2 human intestinal cells. *J. Nutr.* 2006, *136*, 588–594.
- [115] Andre, C. M., Greenwood, J. M., Walker, E. G., Rassam, M. et al., Anti-inflammatory procyanidins and triterpenes in 109 apple varieties. *J. Agric. Food Chem.* 2012, *60*, 10546–10554.
- [116] Bohn, T., Tian, Q., Chitchumroonchokchai, C., Failla, M. L. et al., Supplementation of test meals with fat-free phytosterol products can reduce cholesterol micellarization during simulated digestion and cholesterol accumulation by Caco-2 cells. *J. Agric. Food Chem.* 2007, *55*, 267–272.
- [117] Parker, R. S., Absorption, metabolism, and transport of carotenoids. *FASEB J.* 1996, *10*, 542–551.
- [118] Sy, C., Gleize, B., Dangles, O., Landrier, J. F. et al., Effects of physicochemical properties of carotenoids on their bioaccessibility, intestinal cell uptake, and blood and tissue concentrations. *Mol. Nutr. Food Res.* 2012, *56*, 1385–1397.
- [119] Lesser, S., Cermak, R., Wolfram, S., Bioavailability of quercetin in pigs is influenced by the dietary fat content. *J. Nutr.* 2004, *134*, 1508–1511.
- [120] Chitchumroonchokchai, C., Riedl, K. M., Suksumrarn, S., Clinton, S. K. et al., Xanthones in mangosteen juice are absorbed and partially conjugated by healthy adults. *J. Nutr.* 2012, *142*, 675–680.
- [121] Bohn, T., Blackwood, M., Francis, D., Tian, Q. et al., Bioavailability of phytochemical constituents from a novel soy fortified lycopene rich tomato juice developed for targeted cancer prevention trials. *Nutr. Canc.* 2013, *65*, 919–929.
- [122] Thomson, A. B., Jarocka-Cyrta, E., Faria, J., Wild, G. E., Small bowel review—Part II. *Can. J. Gastroenterol.* 1997, *11*, 159–165.
- [123] Han, X., Shenemail, T., Hongxiang L., Dietary polyphenols and their biological significance. *Int. J. Mol. Sci.* 2007, *8*, 950–988.
- [124] Manach, C., Scalbert, A., Morand, C., Remesy, C. et al., Polyphenols: food sources and bioavailability. *Am. J. Clin. Nutr.* 2004, *79*, 727–747.

- [125] Kern, S. M., Bennett, R. N., Needs, P. W., Mellon, F. A. et al., Characterization of metabolites of hydroxycinnamates in the in vitro model of human small intestinal epithelium caco-2 cells. *J. Agric. Food Chem.* 2003, *51*, 7884–7891.
- [126] Granado-Lorencio, F., Olmedilla-Alonso, B., Herrero-Barbudo, C., Blanco-Navarro, I. et al., In vitro bioaccessibility of carotenoids and tocopherols from fruits and vegetables. *Food Chem.* 2007, *102*, 641–648.
- [127] Cai, K., Bennick, A., Effect of salivary proteins on the transport of tannin and quercetin across intestinal epithelial cells in culture. *Biochem. Pharmacol.* 2006, *72*, 974–980.
- [128] Singh, H., Ye, A., Structural and biochemical factors affecting the digestion of protein-stabilized emulsions. *Curr. Opin. Colloid Interface Sci.* 2013, *18*, 360–370.
- [129] Bitsch, R., Netzel, M., Frank, T., Strass, G. et al., Bioavailability and biokinetics of anthocyanins from red grape juice and red wine. *J. Biomed. Biotechnol.* 2004, *2004*, 293–298.
- [130] Biehler, E., Hoffmann, L., Krause, E., Bohn, T., Divalent minerals decrease micellization and uptake of carotenoids and digestion products into Caco-2 cells. *J. Nutr.* 2011, *141*, 1769–1776.
- [131] Lai, S. K., Wang, Y. Y., Wirtz, D., Hanes, J., Micro- and macrorheology of mucus. *Adv. Drug Deliv. Rev.* 2009, *61*, 86–100.
- [132] Dunnhaupt, S., Barthelmes, J., Hombach, J., Sakloetsakun, D. et al., Distribution of thiolated mucoadhesive nanoparticles on intestinal mucosa. *Int. J. Pharm.* 2011, *408*, 191–199.
- [133] Menard, S., Cerf-Bensussan, N., Heyman, M., Multiple facets of intestinal permeability and epithelial handling of dietary antigens. *Mucosal Immunol* 2010, *3*, 247–259.
- [134] Rogoll, D., Bergmann, H., Hellenschmidt, D., Heinze, J. et al., Influence of apple polyphenols on the intestinal barrier in a colonic cell model. *J. Appl. Bot. Food Qual.* 2010, *83*, 110–117.
- [135] Ulluwishewa, D., Anderson, R. C., McNabb, W. C., Moughan, P. J. et al., Regulation of tight junction permeability by intestinal bacteria and dietary components. *J. Nutr.* 2011, *141*, 769–776.
- [136] Appeldoorn, M. M., Vincken, J. P., Gruppen, H., Hollman, P. C., Procyanidin dimers A1, A2, and B2 are absorbed without conjugation or methylation from the small intestine of rats. *J. Nutr.* 2009, *139*, 1469–1473.
- [137] Shoji, T., Masumoto, S., Moriichi, N., Akiyama, H. et al., Apple procyanidin oligomers absorption in rats after oral administration: analysis of procyanidins in plasma using the porter method and high-performance liquid chromatography/tandem mass spectrometry. *J. Agric. Food Chem.* 2006, *54*, 884–892.
- [138] Chhabra, R. S., Intestinal absorption and metabolism of xenobiotics. *Environ. Health Perspect.* 1979, *33*, 61–69.
- [139] Estudante, M., Morais, J. G., Soveral, G., Benet, L. Z., Intestinal drug transporters: an overview. *Adv. Drug Deliv. Rev.* 2013, *65*, 1340–1356.
- [140] Li, Y., Paxton, J. W., The effects of flavonoids on the ABC transporters: consequences for the pharmacokinetics of substrate drugs. *Exp. Opin. Drug Metab. Toxicol.* 2013, *9*, 267–285.
- [141] Reboul, E., Thap, S., Tourniaire, F., Andre, M. et al., Differential effect of dietary antioxidant classes (carotenoids, polyphenols, vitamins C and E) on lutein absorption. *Br. J. Nutr.* 2007, *97*, 440–446.
- [142] Reboul, E., Absorption of vitamin A and carotenoids by the enterocyte: focus on transport proteins. *Nutrients* 2013, *5*, 3563–3581.
- [143] Schramm, D. D., Karim, M., Schrader, H. R., Holt, R. R. et al., Food effects on the absorption and pharmacokinetics of cocoa flavanols. *Life Sci.* 2003, *73*, 857–869.
- [144] Paine, M. F., Fisher, M. B., Immunochemical identification of UGT isoforms in human small bowel and in caco-2 cell monolayers. *Biochem. Biophys. Res. Commun.* 2000, *273*, 1053–1057.
- [145] Chen, G., Zhang, D., Jing, N., Yin, S. et al., Human gastrointestinal sulfotransferases: identification and distribution. *Toxicol. Appl. Pharmacol.* 2003, *187*, 186–197.
- [146] Shoba, G., Joy, D., Joseph, T., Majeed, M. et al., Influence of piperine on the pharmacokinetics of curcumin in animals and human volunteers. *Planta Med.* 1998, *64*, 353–356.
- [147] Kaulmann, A., Bohn, T., Carotenoids, inflammation and oxidative stress – implications in cellular signalling pathways. *Nutr. Res.* 2014, *34*, 907–929.
- [148] van de Wetering, K., Burkon, A., Feddema, W., Bot, A. et al., Intestinal breast cancer resistance protein (BCRP)/Bcrp1 and multidrug resistance protein 3 (MRP3)/Mrp3 are involved in the pharmacokinetics of resveratrol. *Mol. Pharmacol.* 2009, *75*, 876–885.
- [149] Hussain, M. M., Fatma, S., Pan, X., Iqbal, J., Intestinal lipoprotein assembly. *Curr. Opin. Lipidol.* 2005, *16*, 281–285.
- [150] Escudero-Lopez, B., Calani, L., Fernandez-Pachon, M. S., Ortega, A. et al., Absorption, metabolism, and excretion of fermented orange juice (poly)phenols in rats. *Biofactors* 2014, *40*, 327–335.
- [151] Borges, G., Lean, M. E., Roberts, S. A., Crozier, A., Bioavailability of dietary (poly)phenols: a study with ileostomists to discriminate between absorption in small and large intestine. *Food Funct.* 2013, *4*, 754–762.
- [152] Nardini, M., Cirillo, E., Natella, F., Scaccini, C., Absorption of phenolic acids in humans after coffee consumption. *J. Agric. Food Chem.* 2002, *50*, 5735–5741.
- [153] Natsume, M., Osakabe, N., Oyama, M., Sasaki, M. et al., Structures of (-)-epicatechin glucuronide identified from plasma and urine after oral ingestion of (-)-epicatechin: differences between human and rat. *Free Radic. Biol. Med.* 2003, *34*, 840–849.
- [154] Pimpao, R. C., Dew, T., Figueira, M. E., McDougall, G. J. et al., Urinary metabolite profiling identifies novel colonic metabolites and conjugates of phenolics in healthy volunteers. *Mol. Nutr. Food Res.* 2014, *58*, 1414–1425.
- [155] Felgines, C., Talavera, S., Gonthier, M. P., Texier, O. et al., Strawberry anthocyanins are recovered in urine as glucuro- and sulfoconjugates in humans. *J. Nutr.* 2003, *133*, 1296–1301.

- [156] Lampe, J. W., Isoflavonoid and lignan phytoestrogens as dietary biomarkers. *J. Nutr.* 2003, *133*, 956S–964S.
- [157] Sawai, Y., Kohsaka, K., Nishiyama, Y., Ando, K., Serum concentrations of rutoside metabolites after oral administration of a rutoside formulation to humans. *Arzneimittelforschung* 1987, *37*, 729–732.
- [158] Adlercreutz, H., van der, W. J., Kinzel, J., Attalla, H. et al., Lignan and isoflavonoid conjugates in human urine. *J. Steroid Biochem. Mol. Biol.* 1995, *52*, 97–103.
- [159] Axelson, M., Setchell, K. D., The excretion of lignans in rats – evidence for an intestinal bacterial source for this new group of compounds. *FEBS Lett.* 1981, *123*, 337–342.
- [160] Duenas, M., Surco-Laos, F., Gonzalez-Manzano, S., Gonzalez-Paramas, A. M. et al., Deglycosylation is a key step in biotransformation and lifespan effects of quercetin-3-O-glucoside in *Caenorhabditis elegans*. *Pharmacol. Res.* 2013, *76*, 41–48.
- [161] Aura, A. M., O’Leary, K. A., Williamson, G., Ojala, M. et al., Quercetin derivatives are deconjugated and converted to hydroxyphenylacetic acids but not methylated by human fecal flora in vitro. *J. Agric. Food Chem.* 2002, *50*, 1725–1730.
- [162] Crozier, A., Absorption, metabolism, and excretion of (-)-epicatechin in humans: an evaluation of recent findings. *Am. J. Clin. Nutr.* 2013, *98*, 861–862.
- [163] Actis-Goretta, L., Leveques, A., Rein, M., Teml, A. et al., Intestinal absorption, metabolism, and excretion of (-)-epicatechin in healthy humans assessed by using an intestinal perfusion technique. *Am. J. Clin. Nutr.* 2013, *98*, 924–933.
- [164] Clifford, M. N., van der Hooft, J. J., Crozier, A., Human studies on the absorption, distribution, metabolism, and excretion of tea polyphenols. *Am. J. Clin. Nutr.* 2013, *98*, 1619S–1630S.
- [165] Monagas, M., Urpi-Sarda, M., Sanchez-Patan, F., Llorach, R. et al., Insights into the metabolism and microbial biotransformation of dietary flavan-3-ols and the bioactivity of their metabolites. *Food Funct.* 2010, *1*, 233–253.
- [166] Kuijsten, A., Arts, I. C., Vree, T. B., Hollman, P. C., Pharmacokinetics of enterolignans in healthy men and women consuming a single dose of secoisolariciresinol diglucoside. *J. Nutr.* 2005, *135*, 795–801.
- [167] Seeram, N. P., Henning, S. M., Zhang, Y., Suchard, M. et al., Pomegranate juice ellagitannin metabolites are present in human plasma and some persist in urine for up to 48 hours. *J. Nutr.* 2006, *136*, 2481–2485.
- [168] Gross, M., Pfeiffer, M., Martini, M., Campbell, D. et al., The quantitation of metabolites of quercetin flavonols in human urine. *Cancer Epidemiol. Biomarkers Prev.* 1996, *5*, 711–720.
- [169] Vetrani, C., Rivellese, A. A., Annuzzi, G., Mattila, I. et al., Phenolic metabolites as compliance biomarker for polyphenol intake in a randomized controlled human intervention. *Food Res. Int.* 2014, *63*, 233–238.
- [170] Griffith, L. G., Wells, A., Stolz, D. B., Engineering liver. *Hepatology* 2014, *60*, 1426–1434.
- [171] Mandenius, C. F., Andersson, T. B., Alves, P. M., Batzl-Hartmann, C. et al., Toward preclinical predictive drug testing for metabolism and hepatotoxicity by using in vitro models derived from human embryonic stem cells and human cell lines – a report on the Vitrocellomics EU-project. *Altern. Lab. Anim.* 2011, *39*, 147–171.
- [172] Barron, D., Smarrito-Menzozi, C., Viton, F., (Bio)chemical labelling tools for studying absorption & metabolism of dietary phenols – an overview. *Curr. Org. Chem.* 2012, *16*, 663–690.
- [173] Saha, S., Hollands, W., Needs, P. W., Ostertag, L. M. et al., Human O-sulfated metabolites of (-)-epicatechin and methyl(-)-epicatechin are poor substrates for commercial aryl-sulfatases: implications for studies concerned with quantifying epicatechin bioavailability. *Pharmacol. Res.* 2012, *65*, 592–602.
- [174] Actis-Goretta, L., Leveques, A., Giuffrida, F., Romanov-Michailidis, F. et al., Elucidation of (-)-epicatechin metabolites after ingestion of chocolate by healthy humans. *Free Radic. Biol. Med.* 2012, *53*, 787–795.
- [175] Jacobs, D. M., Deltimple, N., van, V. E., van Dorsten, F. A. et al., (1)H NMR metabolite profiling of feces as a tool to assess the impact of nutrition on the human microbiome. *NMR Biomed.* 2008, *21*, 615–626.
- [176] Hanhineva, K., Aura, A. M., Rogachev, I., Matero, S. et al., In vitro microbiotic fermentation causes an extensive metabolite turnover of rye bran phytochemicals. *PLoS One* 2012, *7*, e39322.
- [177] Grun, C. H., van Dorsten, F. A., Jacobs, D. M., Le, B. M. et al., GC-MS methods for metabolic profiling of microbial fermentation products of dietary polyphenols in human and in vitro intervention studies. *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* 2008, *871*, 212–219.
- [178] Oleaga, C., Ciudad, C. J., Izquierdo-Pulido, M., Noe, V., Cocoa flavanol metabolites activate HNF-3beta, Sp1, and NFY-mediated transcription of apolipoprotein AI in human cells. *Mol. Nutr. Food Res.* 2013, *57*, 986–995.
- [179] Awad, A. B., Fink, C. S., Phytosterols as anticancer dietary components: evidence and mechanism of action. *J. Nutr.* 2000, *130*, 2127–2130.
- [180] Wong, A., Chemical and microbiological considerations of phytosterols and their relative efficacies in functional foods for the lowering of serum cholesterol levels in humans: a review. *J. Funct. Foods* 2014, *6*, 60–72.
- [181] Moussa, M., Landrier, J. F., Reboul, E., Ghiringhelli, O. et al., Lycopene absorption in human intestinal cells and in mice involves scavenger receptor class B type I but not Niemann-Pick C1-like 1. *J. Nutr.* 2008, *138*, 1432–1436.
- [182] Sen, A., Ren, J., Ruffin, M. T., Turgeon, D. K. et al., Relationships between serum and colon concentrations of carotenoids and fatty acids in randomized dietary intervention trial. *Cancer Prev. Res. (Phila)* 2013, *6*, 558–565.
- [183] Khachik, F., Carvalho, L., Bernstein, P. S., Muir, G. J. et al., Chemistry, distribution, and metabolism of tomato carotenoids and their impact on human health. *Exp. Biol. Med. (Maywood)* 2002, *227*, 845–851.

- [184] Faria, A., Meireles, M., Fernandes, I., Santos-Buelga, C. et al., Flavonoid metabolites transport across a human BBB model. *Food Chem.* 2014, *149*, 190–196.
- [185] Sergent, T., Garsou, S., Schaut, A., De, S. S. et al., Differential modulation of ochratoxin A absorption across Caco-2 cells by dietary polyphenols, used at realistic intestinal concentrations. *Toxicol. Lett.* 2005, *159*, 60–70.
- [186] Bailey, D. G., Malcolm, J., Arnold, O., Spence, J. D., Grapefruit juice–drug interactions. *Br. J. Clin. Pharmacol.* 1998, *46*, 101–110.
- [187] Lampe, J. W., Interindividual differences in response to plant-based diets: implications for cancer risk. *Am. J. Clin. Nutr.* 2009, *89*, 1553S–1557S.
- [188] Yum, K., Hong, S. G., Healy, K. E., Lee, L. P., Physiologically relevant organs on chips. *Biotechnol. J.* 2014, *9*, 16–27.
- [189] Surco-Laos, F., Cabello, J., Gomez-Orte, E., Gonzalez-Manzano, S. et al., Effects of O-methylated metabolites of quercetin on oxidative stress, thermotolerance, lifespan and bioavailability on *Caenorhabditis elegans*. *Food Funct.* 2011, *2*, 445–456.
- [190] Cilla, A., Alegria, A., Barbera, R., in: Morales-Gonzales, J. A. (Ed.), *Oxidative Stress and Chronic Degenerative Diseases – a Role for Antioxidants Rijeka*, InTech, Croatia 2013, 131–151.
- [191] Tavares, L., Figueira, I., Macedo, D., McDougall, G. J. et al., Neuroprotective effect of blackberry (*Rubus* sp.) polyphenols is potentiated after simulated gastrointestinal digestion. *Food Chem.* 2012, *131*, 1443–1452.
- [192] Tavares, L., Figueira, I., McDougall, G. J., Vieira, H. L. et al., Neuroprotective effects of digested polyphenols from wild blackberry species. *Eur. J. Nutr.* 2013, *52*, 225–236.
- [193] Ferruzzi, M. G., The influence of beverage composition on delivery of phenolic compounds from coffee and tea. *Physiol. Behav.* 2010, *100*, 33–41.
- [194] Konishi, Y., Shimizu, M., Transepithelial transport of ferulic acid by monocarboxylic acid transporter in Caco-2 cell monolayers. *Biosci. Biotechnol. Biochem.* 2003, *67*, 856–862.
- [195] Watanabe, H., Yashiro, T., Tohjo, Y., Konishi, Y., Non-involvement of the human monocarboxylic acid transporter 1 (MCT1) in the transport of phenolic acid. *Biosci. Biotechnol. Biochem.* 2006, *70*, 1928–1933.
- [196] Gill, R. K., Saksena, S., Alrefai, W. A., Sarwar, Z. et al., Expression and membrane localization of MCT isoforms along the length of the human intestine. *Am. J. Physiol. Cell Physiol.* 2005, *289*, C846–C852.
- [197] Wolfram, S., Block, M., Ader, P., Quercetin-3-glucoside is transported by the glucose carrier SGLT1 across the brush border membrane of rat small intestine. *J. Nutr.* 2002, *132*, 630–635.
- [198] Reboul, E., Borel, P., Proteins involved in uptake, intracellular transport and basolateral secretion of fat-soluble vitamins and carotenoids by mammalian enterocytes. *Prog. Lipid Res.* 2011, *50*, 388–402.