# Kinetics of Microbial Translocation Markers in Patients on Efavirenz or Lopinavir/r Based Antiretroviral Therapy

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# Abstract

**Objectives:** We investigated whether there are differences in the effects on microbial translocation (MT) and enterocyte damage by different antiretroviral therapy (ART) regimens after 1.5 years and whether antibiotic use has impact on MT. In a randomized clinical trial (NCT01445223) on first line ART, patients started either lopinavir/r (LPV/r) (n = 34) or efavirenz (EFV) containing ART (n = 37). Lipopolysaccharide (LPS), sCD14, anti-flagellin antibodies and intestinal fatty acid binding protein (I-FABP) levels were determined in plasma at baseline (BL) and week 72 (w72).

**Results:** The levels of LPS and sCD14 were reduced from BL to w72 (157.5 pg/ml vs. 140.0 pg/ml, p = 0.0003; 3.13 ug/ml vs. 2.85 ug/ml, p = 0.005, respectively). The levels of anti-flagellin antibodies had decreased at w72 (0.35 vs 0.31 [OD]; p<0.0004), although significantly only in the LPV/r arm. I-FABP levels increased at w72 (2.26 ng/ml vs 3.13 ng/ml; p<0.0001), although significantly in EFV treated patients only. Patients given antibiotics at BL had lower sCD14 levels at w72 as revealed by ANCOVA compared to those who did not receive ( $\Delta = -0.47 \mu g/ml$ ; p = 0.015).

**Conclusions:** Markers of MT and enterocyte damage are elevated in untreated HIV-1 infected patients. Long-term ART reduces the levels, except for I-FABP which role as a marker of MT is questionable in ART-experienced patients. Why the enterocyte damage seems to persist remains to be established. Also antibiotic usage may influence the kinetics of the markers of MT.

Trial Registration: ClinicalTrials.gov NCT01445223 http://clinicaltrials.gov/ct2/show/NCT01445223?term = NCT01445223

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### Introduction

A sustained control of the human immunodeficiency virus type 1 (HIV-1) replication is obtained by antiretroviral therapy (ART) in the majority of patients, reducing plasma HIV-1 load to undetectable levels. However, HIV-1 persists in reservoirs like latently infected CD4+ T cells [1,2] and in body compartments that have restricted permission for antiretroviral drugs. It is hypothesized that these reservoirs are refilled by the low grade viral replication seen in patients on suppressive ART who otherwise have undetectable viremia by routine assays [3,4]. Translocation of bacterial products across a damaged gut-blood barrier has been proposed to be one important mechanism for the persistence of a chronic immune activation which is found also in well-treated patients [5,6]. There is growing evidence that this

immune activation may contribute to the low-grade viremia and e.g. cardiovascular and CNS complications [7,8,9].

Several markers are used to assess microbial translocation (MT) in patients with HIV or inflammatory bowel disease [10], such as microbial products [lipopolysaccharide (LPS), plasma bacterial 16SrDNA, anti-flagellin antibodies] [11], markers of the systemic response to bacterial products (sCD14, LPS-binding protein), and of enterocyte damage [intestinal fatty acid binding protein (I-FABP)] [12,13]. During successful ART, MT and systemic immune activation are usually reduced, but not normalized, suggesting that the damage of the gut-blood barrier is only partly restored [5]. The reasons why this improvement varies between patients are not known.

The origin of low-level viremia in patients on suppressive ART has been disputed. Residual virus replication from anatomical

compartments like the gut could be one of the explanations. Firstly, levels of HIV-1 DNA and RNA were substantially elevated in the gut compared with peripheral blood in patients on ART with <40 HIV-1 RNA copies/ml, indicating that the gut may serve as a potential source of viremia during suppressive ART [14]. Additionally, in a set of patients on long-term ART, levels of HIV DNA in sigmoid colon were positively correlated to plasma LPS levels [15], suggesting a connection between residual viremia and MT. In a cross-sectional study of patients with persistently undetectable HIV-1 RNA, a higher proportion of participants treated with nevirapine and efavirenz achieved <2.5 HIV-1 RNA copies/ml compared to lopinavir/r based ART [16]. Given the link between MT and low-level viremia, we assumed that choice of ART could differently affect kinetics of MT markers.

In the present study, we analyzed the levels of LPS, sCD14, I-FABP, and anti-flagellin antibodies, at baseline (BL) and after 72 weeks (w72) of ART in a controlled randomized clinical trial in which the patients received either lopinavir/r +2 nucleoside analogues (NRTI) or efavirenz +2 NRTI. Additionally, we studied if ongoing antibiotics treatment had an impact on the explored parameters.

### Methods

### Subjects

During 2004-2007, 239 HIV-1 infected subjects received allocated intervention after written consent in a Scandinavian randomized clinical phase IV efficacy trial (RCT) (Clinical Trials.gov identifier: NCT01445223). The protocol for this trial and supporting CONSORT checklist are available as supporting information; see Checklist S1 and Protocol S1. The study protocol was approved by the Regional Ethics Committee (Gothenburg Ö739-03). The study design and participants have been described elsewhere [17,18]. In our substudy, the patients were randomized to receive either efavirenz (EFV) +2 NRTI once daily (n = 37) or ritonavir-boosted lopinavir (LPV/r) +2 NRTI twice daily (n = 34) (Figure 1). Totally 59 patients were excluded from our analysis because of insufficient remaining plasma volumes after the main study analyzes. CD4+ T-cell count, viral load (VL), rate of hepatitis B/C co-infection and age were similar in the group of excluded patients at BL, compared to the substudy group.

Data on antibiotic therapy was available in 63 patients of whom 29 were given antibiotics at baseline (BL) (n = 27) and/or week72 (w72) (n = 10), while 34 had not received antibiotics at any of the two time points (Table 1). At BL, the patients received: cotrimoxazole (TMP-SMX) as *Pneumocystis jiroveci* prophylaxis (PCP) (n = 24) or for treatment of pneumonia (n = 2), or clindamycin (n = 1). Also, two were on *Mycobacterium tuberculosis* treatment, and one was on fluconazole. At w72, TMP-SMX was given as PCP prophylaxis (n = 9), and one patient received nitrofurantoin; 8 of these 10 had had TMP-SMX at baseline.

### CD4<sup>+</sup> T-cells and Plasma HIV-1 RNA

HIV RNA load and CD4<sup>+</sup> T-cell counts were measured as part of the clinical routine with Cobas Amplicor (Roche Molecular Systems Inc., Branchburg, New Jersey, USA), and flow cytometry, respectively.

### Microbial Translocation Markers

Plasma samples obtained at the sampling day were frozen at  $-80^{\circ}$ C and later thawed. The analyses were performed blindly in relation to clinical data and treatment outcome. Levels of LPS were measured by limulus amebocyte assay (LAL, Lonza, Maryland, USA) as previously described [5]. sCD14 and I-FABP

levels were determined by enzyme-linked immunosorbent assays (R&D Systems, USA and DS Pharma Biomedical Co, Japan; respectively), according to the manufacturers instructions [19]. Samples at BL and w72 from the same patient were assayed on the same plate.

Antibody titers to flagellin, and total IgG levels were assessed by an in-house anti-flagellin specific IgG ELISA [20] using purified flagellin monomers from *S. typhimurium* (InvivoGen, USA). It is known that human sera have a similar recognition pattern of flagellin monomers whether isolated from flagellated *E. coli* or *S. typhimurium* [21]. Briefly, microwell plates (MWP) were coated overnight with purified flagellin from *S. typhimurium* (25 ng/well). The following day, plasma samples from HIV-1 patients were diluted 1:1000 and applied to the MWP. After incubation and washing, the MWPs were incubated with HRP-conjugated antihuman IgG. For total IgG ELISA, the manufacturer's procedure was followed (MABTECH, Nacka, Sweden).

### Statistical Analysis

Data were analyzed using GraphPad Prism v. 5.02 and R 2.13.1. Independent groups were compared using Mann-Whitney U-test and paired data analyzed with Wilcoxon signed rank test. Correlations were analyzed by Spearmans rank test. Differences in LPS, sCD14, I-FABP and anti-flagellin IgG levels between patients with or without antibiotics were analyzed with ANCOVA using covariates age, sex, log viral load and CD4+ T-cell count at BL or w72, and residual plots were inspected. Model selection was done by backward elimination with removal of variables if P>0.1.

### Results

#### HIV RNA Load and Recovery of CD4+ T-cells

Following initiation of ART, all patients achieved plasma HIV-RNA <50 copies/mL within 24 weeks, except for five patients (all <250 c/mL). Four of these reached undetectable HIV-RNA at w72. Two patients had a relapse with detectable HIV-RNA at w72 (190 respective 640 c/mL). No statistically significant differences were found at BL or follow-up between the two arms (Table 2). BL CD4+ T-cells in the LPV/r group (median 155 [IQR 119.5–203.3] cells/ul) and in the EFV group (130 [IQR 69–200] cells/ul) did not differ significantly (Table 2). There was no difference with regard to CD4+ T-cell recovery at w72 (LPV/r: 199 [IQR 135–294]; EFV: 156 [IQR 90–270]) or w48 (p = 0.27) (data not shown). At w72 the CD4+ T-cell count tended to be higher in the LPV/r group as compared to the EFV group (data not shown). There was no correlation between change in CD4+ T-cells and change of any of the markers between BL and w72.

## Decrease of LPS and sCD14 after 72 Weeks of ART

The overall plasma levels of LPS were reduced at w72 compared to BL (157.5 pg/ml [IQR 128.2–171.1] vs. 140.0 [IQR 127.8–155.2]; p = 0.0003) (Figure 2). The reduction in LPS levels and absolute change between BL and w72 was similar in both treatment arms (data not shown). No significant correlation was observed between LPS and CD4+ T-cell count, VL, sCD14 or I-FABP, respectively.

Similar to LPS, plasma levels of sCD14 were reduced at w72 compared to BL (3.13 ug/ml [IQR 2.7–3.8] vs. 2.85 ug/ml [IQR 2.4–3.4]; p = 0.005) (Figure 3) in both treatment arms. The CD4+ T-cell count and sCD14 levels showed a highly significant negative correlation at BL ( $\rho = -0.42$ , p = 0.0003) (Figure 4A) and at w72 ( $\rho = -0.32$ , p = 0.007). Viral load and sCD14 levels correlated at baseline ( $\rho = 0.42$ , p = 0.0002) (Figure 4B).



# Figure 1. Trial profile and reasons for discontinuations in each group. doi:10.1371/journal.pone.0055038.g001

## Elevated Levels of I-FABP after 72 Weeks of EFV Containing ART

At BL, no difference was found in plasma I-FABP levels between the treatment groups. At w72, the overall I-FABP concentrations had increased (median 2.26 ng/ml [IQR 1.4–3.6] at BL vs 3.13 ng/ml [IQR 1.8–4.9] at w72; p<0.0001) (Figure 5). The elevation was significant only in the EFV arm (2.32 [IQR 1.5–3.8] vs. 4.29 [IQR 2.4–5.9]; p<0.0001), but not in the LPV/r arm (2.19 [IQR 1.3–3.4] vs. 2.56 [IQR 1.7–3.5]). The difference in elevation between the two arms was significant (p = 0.035). A significant difference was also found between the treatment arms at w72 (EFV: median 4.29 [IQR 2.4–5.9]; LPV/r: 2.56 [IQR 1.7–3.5]; p = 0.003). There was no significant correlation between I-FABP levels and CD4+ T-cells, VL, and sCD14, respectively.

# Decrease of Anti-flagellin Antibodies after 72 Weeks of ART

Anti-flagellin antibodies were detected in all subjects. ART reduced the levels in the whole cohort (median OD 0.35 [IQR

0.22–0.61] at BL vs. 0.31 [IQR 0.21–0.5]) at w72; p=0.0004) (Figure 6). In the LPV/r arm, anti-flagellin antibodies decreased significantly (median OD 0.34 [IQR 0.22–0.5] vs. 0.25 [IQR 0.14–0.49]; p=0.001), but not in the EFV treated patients (median OD 0.35 [IQR 0.23–0.63] vs. 0.37 [IQR 0.24–0.51]; p=0.06).

As expected, the total IgG concentrations were reduced at w72 (median OD 1.02 [IQR 0.99–1.07] at BL vs 0.99 [IQR 0.96–1.02] at w72; p = 0.0001) in the entire cohort, and in both treatment arms (LPV/r arm median OD 1.02 [IQR 0.99–1.05] at BL vs 0.98 [IQR 0.94–1.01] at w72; p < 0.0001); (EFV arm median OD 1.03 [IQR 0.98–1.08] at BL vs 1 [IQR 0.96–1.03] at w72; p < 0.0001). The anti-flagellin IgG:total IgG ratio was reduced between BL and w72 in whole cohort (median 0.34 [IQR 0.22–0.53] vs 0.3 [IQR 0.21–0.5]; p = 0.005) and LPV/r arm (median 0.34 [IQR 0.22–0.48] vs 0.26 [IQR 0.15–0.47]; p = 0.003), supporting that the decrease of anti-flagellin IgG was specific. This reduction was not observed in EFV treated (median 0.34 [IQR 0.23–0.59] vs 0.37 [IQR 0.25–0.52]; p = 0.31). LPS and anti-flagellin antibodies correlated at BL

Table 1. Baseline characteristics of antibiotic treated patients.

	All	Antibiotic treated	Other
Subjects [number]	63	27	36
CD4+ count/ul [median (range)]	160 (90–205)	120 (30–150)	180 (138–220)
HIV-1 RNA [median log10 copies/ml (range)]	5.25 (2.95–7.07)	5.34 (3.86–7.07)	5.15 (2.95-6.66)

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Table 2. Characteristics of HIV-1 infected patients.

	All	Lopinavir-arm*	Efavirenz-arm*
Subjects [number]	71	34	37
Age [years, median (range)]	37 (19–70)	41 (19–68)	35 (20–70)
Sex [male/female]	51/20	24/10	27/10
Hepatitis [B/C]	1/3	1/2	0/1
CD4+ count/ul [median (range)]			
BL	140 (0–370)	155 (11–370)	130 (0–270)
w72	330 (73–1260)	370 (160–1260)	300 (73–843)
HIV-1 RNA [median log10copies/ml (range)]			
BL	5.3 (2.95–7.07)	5.3 (2.95–6.72)	5.3 (3.23–7.07)
w72	0 (0–2.81)	0 (0–2.28)	0 (0–2.81)
Antibiotics use <sup>1</sup>			
BL	27	10	17
w72	10	3	7

\*There were no statistically significant differences in age, CD4+ -cell count or HIV-1 RNA between the two treatment arms at BL. Abbreviations: BL, baseline. <sup>1</sup>Data from 63 subjects were available.

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 $(\rho = 0.25, p = 0.04)$  and at w72  $(\rho = 0.31, p = 0.01)$ . On the contrary, there was no significant correlation at BL or w72 between anti-flagellin antibodies and CD4+ T-cells, VL, sCD14 or I-FABP, respectively.

# Antibiotic Treatment and Markers of Microbial Translocation

We explored the impact of antibiotics on markers of MT with three comparisons using ANCOVA adjusting for significant



**Figure 2. Levels of LPS.** Levels of LPS were significantly reduced after 72 weeks of antiretroviral therapy in the whole cohort, with a unified pattern in both treatment arms. Data are shown as individual values with corresponding baseline LPS levels at X-axis and week 72 levels at Y-axis, respectively. Individuals treated with lopinavir (LPV/r) are indicated with shaded and efavirenz (EFV) with black circles. Dotted lines refer to median values at the corresponding time-point. doi:10.1371/journal.pone.0055038.g002



**Figure 3. Levels of sCD14.** Levels of sCD14 were significantly reduced after 72 weeks of antiretroviral therapy in the whole group, with a unified pattern in both treatment arms. Data are shown as individual values with corresponding baseline sCD14 levels at X-axis and week 72 levels at Y-axis, respectively. Individuals treated with lopinavir (LPV/r) are indicated with shaded and efavirenz (EFV) with black circles. Dotted lines refer to median values at the corresponding time-point. doi:10.1371/journal.pone.0055038.g003

covariates. At BL, no difference was found between patients with or without antibiotics. At w72, the levels of sCD14 were lower in patients with ongoing antibiotic treatment vs. those without ( $\Delta = -0.70 \ \mu g/ml$ , p = 0.017). Also, in the untreated group at w72 the patients who had been treated with antibiotics at baseline had lower levels than those who had been untreated at baseline ( $\Delta = -0.47 \ \mu g/ml$ ; p = 0.015). Finally, the change in the sCD14 levels between BL and w72 was calculated. A difference between

antibiotic treated vs untreated patients at BL was then found (median  $\Delta = -0.85$  vs  $-0.08 \ \mu g/ml$ , p = 0.02) (Figure 7). No effect of antibiotics was observed on the levels of LPS, I-FABP and anti-flagellin antibodies, respectively.

### Discussion

The microbial translocation (MT) markers assessed in our study (LPS, sCD14, I-FABP, anti-flagellin antibodies) were all increased



Figure 4. Correlations between sCD14, CD4+ T-cells and HIV-1 RNA. At baseline, significant correlations were found between sCD14 and CD4-T cell counts (A) or HIV-1 RNA viral load (B), respectively. Rho and p-value refer to Spearman's rank test. doi:10.1371/journal.pone.0055038.g004



**Figure 5. Levels of Intestinal Fatty Acid Binding Protein (I-FABP).** Plasma levels of I-FABP were significantly elevated after 72 weeks of antiretroviral treatment as compared to baseline in the whole cohort. Stratified to treatment arms, efavirenz (EFV) treated patients had significantly increased levels, but not the patients on lopinavir/r (LPV/r) therapy. Data are shown as individual values with corresponding baseline I-FABP levels at X-axis and week 72 levels at Y-axis, respectively. Individuals treated with lopinavir (LPV/r) are indicated with shaded and efavirenz (EFV) with black circles. Dotted lines refer to median values at the corresponding time-point. doi:10.1371/journal.pone.0055038.g005

in plasma before treatment and ART influenced the levels during the follow-up period of 72 weeks. The reliability of our results is strengthened by the fact that the patients were included in a randomized clinical trial and were followed closely before, during and after the study. Based on the randomization, we found that the effects on the selected MT parameters could partly differ depending on the type of ART and if antibiotics were given or not. We therefore suggest that other factors than HIV treatment in itself may be relevant for the kinetics of the biomarkers which should be considered when interpreting studies on MT in HIV-1 infected patients.

Data from our RCT cohort showed that the plasma levels of LPS were significantly reduced after 72 weeks of ART with similar pattern in patients treated with NNRTI or PI/r containing therapy. Similar observations on the decrease of plasma LPS after ART have been reported in some [5,6,22], but not all studies [23,24]. Although, estimating MT by measurement of plasma LPS levels is common, both LAL assay variability as well as patient cohort heterogeneity (e.g. diverse rate of opportunistic conditions or hepatitis co-infection) may lie behind the different results. E.g., in a recent work, the kinetics of MT markers (measured by LPS and sCD14 levels) differed according to the severity of pre-ART CD4+ T-cell count [25].

The effect on sCD14 has also been disputed with studies showing decline [23,26] or increase [22,24] after initiation of ART. In our study, the sCD14 levels were reduced during ART with a unified pattern in the two treatment arms which is in line with a decreased MT during 72 weeks of ART. It has also been suggested that sCD14 independently predicts mortality in HIV-1 infected patients when adjusting for other key markers [13]. We found a strong inverse correlation between CD4+ T-cells and sCD14 as well as a direct correlation between sCD14 and VL [27]. These findings warrant the hypothesis that higher viral replication is associated with more profound inflammatory response (reflected by sCD14) and leads to lower CD4 T cell counts that also reflect poorer surveillance of gut microbes. The viscous circle of viral replication and inflammation is created and maintained by the "leaky gut" as suggested by Douek et al [28]. One should acknowledge that the sCD14 levels are also elevated in other infections (RSV, Dengue Virus, Mycobacteria) and inflammatory conditions [29,30]. Thus, although analysis of sCD14 is important for the evaluation of MT in the setting of HIV-1 infected patients, it cannot be considered as a specific marker for MT alone.

Our untreated patients exhibited elevated plasma levels of I-FABP, as compared to historical healthy controls [19], which is in line with the suggestion that I-FABP can be applied as a marker of enterocyte damage and possibly of MT in HIV-1 infected patients [19]. Actually, the levels were similar to those reported in inflammatory bowel disease (IBD) [31], supporting the presence of a significant damage to the enterocytes in HIV-1 patients, at least in those who have a relatively advanced immunodeficiency. To our knowledge, no data on the effect of ART on I-FABP levels have earlier been published. Surprisingly, we found that the I-FABP levels increased in the whole cohort despite 72 weeks of efficient ART. A subgroup analysis revealed that the I-FABP increase occurred in the patients treated with EFV, but not in



**Figure 6. Plasma levels of anti-flagellin antibodies.** A significant reduction of anti-flagellin IgG levels was seen after 72 weeks of antiretroviral treatment compared to baseline in the whole cohort. Plasma levels of anti-flagellin IgG decreased significantly in lopinavir/r (LPV/r) treated patients, but not during efavirenz (EFV) therapy. Data are shown as individual values with corresponding baseline anti-flagellin IgG levels at X-axis and week 72 levels at Y-axis, respectively. Individuals treated with lopinavir (LPV/r) are indicated with shaded and efavirenz (EFV) with black circles. Dotted lines refer to median values at the corresponding time-point. doi:10.1371/journal.pone.0055038.g006

those with LPV/r. This difference between the two treatment arms could not be explained by any coexisting liver failure, gastrointestinal disease (Inflammatory Bowel Disease, gastroenteritis) or reported side effects such as diarrhea (data not shown). The finding was unexpected particularly in view of the reports claiming a lower level of the residual viremia in EFV-treated patients as



**Figure 7. Change (** $\Delta$ **) in sCD14 levels in relation to antibiotics.** Change ( $\Delta$ ) in sCD14 levels between baseline (BL) and 72 weeks of treatment (w72). Patients treated with antibiotics at BL had a larger reduction in sCD14 levels at w72 than those who did not receive antibiotics at BL. P-value refers to Mann-Whitney test. doi:10.1371/journal.pone.0055038.g007

compared to those on a PI/r regimen [16,32,33]. Although I-FABP has been firmly described as marker of enterocyte damage in other diseases [34,35], we cannot exclude that the increased I-FABP concentrations in our patients might be related to other events such as drug toxicity or metabolic lipid changes. However, the available laboratory blood and lipid-profile data (baseline and follow up) from our patients did not support these relationships (data not shown). Another hypothesis could be based on that EFV may induce oxidative stress related cell apoptosis [36,37]. Conversely, PIs are attributed the anti-apoptotic effect [38,39]. The CD4+ T-cell recovery did not differ significantly between treatment arms, but tended to be higher in LPV/r group. As such, the possibility of immune reconstitution in the gut as a cause of the increasing I-FABP levels in EFV group thus seems to be less likely. Further studies are required to determine if the difference reported by us is related to variation in apoptosis or enhanced turnover of intestinal epithelial cells and as a consequence shedding of I-FABP. In the light of our report, the use of I-FABP for evaluating MT in patients introduced to ART seems questionable, as the kinetics differed compared to the other markers of MT. Most likely the systemic I-FABP levels in patients on ART do not reflect only MT itself. Further studies that include also intestinal biopsies should address these questions.

The bacterial flagellin is known as a microbial compound with strong immune modulatory properties that has an essential role in conditions with intestinal damage, like IBD. Hence, flagellin is regarded as a dominant immune antigen in Crohns disease (CD) where the antibodies to bacterial flagellin (Anti-CBir1) are detected in about half of patients [40]. Recently we found elevated levels of flagellin-specific IgG in three cohorts of HIV-1 infected patients [11]. Additionally, two years of ART reduced the levels of anti-flagellin IgG, although they were not normalized [20]. In the present study, we confirm and expand our previous observations. Thus, the baseline anti-flagellin levels were increased in all patients and decreased significantly after 72 weeks of ART. However, stratifying the cohort into the treatment arms showed that the significant decrease occurred in the LPV/r containing arm only. This effect was not due to the general polyclonal B-cells activation as the normalization of specific anti-flagellin antibodies to total IgG levels yielded similar results. Our hypothesis is therefore that the decline of the anti-flagellin antibodies was due to decrease of exposure for flagellin itself, as a consequence of restoration of the gut-blood barrier. We also claim that the positive correlation between anti-flagellin antibodies and LPS supports the credibility of anti-flagellin antibodies as a marker of MT, theoretically being more specific than e.g. sCD14, which is a polyclonal marker in nature [41]. The relevance of the observed difference between treatment arms is not clear. It is thus possible, that since both arms tended to decrease the levels of anti-flagellin antibodies, differences in time kinetics between the arms could possibly result in a faster decline in LPV/r group.

Antibiotic treatment leads to changes in the gut microbiota [42,43]. Also, plasma LPS levels are reduced in SIV infected

## References

- Shen L, Siliciano RF (2008) Viral reservoirs, residual viremia, and the potential of highly active antiretroviral therapy to eradicate HIV infection. J Allergy Clin Immunol 122: 22–28.
- Karlsson AC, Gaines H, Sallberg M, Lindback S, Sonnerborg A (1999) Reappearance of founder virus sequence in human immunodeficiency virus type l-infected patients. J Virol 73: 6191–6196.
- Palmer S, Maldarelli F, Wiegand A, Bernstein B, Hanna GJ, et al. (2008) Lowlevel viremia persists for at least 7 years in patients on suppressive antiretroviral therapy. Proc Natl Acad Sci U S A 105: 3879–3884.
- Lindkvist A, Eden A, Norstrom MM, Gonzalez VD, Nilsson S, et al. (2009) Reduction of the HIV-1 reservoir in resting CD4+ T-lymphocytes by high

macaques that are treated with "gut sterilizing" antibiotics [6]. However, so far the impact of antibiotics has not been considered in studies on MT in HIV-1 infected patients. In our study, antibiotic treatment was found to influence the sCD14 levels. ANCOVA-analysis thus revealed that former use of antibiotics at baseline as well as ongoing antibiotic treatment lowered sCD14 levels at week 72 compared to non-antibiotic treated individuals. Additionally, the group using antibiotics at base-line had a larger decrease in sCD14 levels up to week 72. A higher reduction of sCD14 levels in antibiotic treated patients could be due to decreased LPS signaling as a consequence of altered gramnegative intestinal flora. The use of TMP-SMX prophylaxis in ART naïve patients has been associated with a lower annual increase in HIV-1 RNA [44]. Antibiotics like minocycline decrease monocyte activation and can dampen the general inflammation as shown in SIV model [45]. Moreover, rifaximin, a broad-spectrum antibiotic with minimal systematic uptake, has recently been shown to affect endotoxinemia in patients with decompensated cirrhosis. Thus, plasma LPS levels were significantly reduced after 8 weeks course of this antibiotic [46]. These findings together suggest that antibiotic treatment may influence levels of MT markers, potentially providing a bias in studies not compensating for the use.

Collectively, we found that markers of MT were reduced after 72 weeks of ART. EFV and LPV/r based ART, which had similar virological outcome, presented with somewhat different effects on the kinetics of MT markers. Thus, although the profiles of the established markers LPS and sCD14 were unified after 72 weeks of treatment, the levels of I-FABP increased and anti-flagellin antibodies were not significantly reduced, respectively, in the patients on EFV containing therapy. Our data underscore the importance of unraveling the multifaceted correlations between microbial translocation, immune activation, antibiotics usage, HIV-1 replication and the chosen ART. Further longitudinal studies should address this complex issue.

### Supporting Information

# Checklist S1 CONSORT Checklist.

**Protocol S1 Trial Protocol.** (DOC)

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### **Author Contributions**

Conceived and designed the experiments: JV, PN, AS, HF, TK, L-MA, MG. Performed the experiments:. BB, SA, JN. Analyzed the data: JV, PN, AS. Wrote the paper: JV, PN, AS.

dosage intravenous immunoglobulin treatment: a proof-of-concept study. AIDS Res Ther 6: 15.

- Troseid M, Nowak P, Nystrom J, Lindkvist A, Abdurahman S, et al. (2010) Elevated plasma levels of lipopolysaccharide and high mobility group box-1 protein are associated with high viral load in HIV-1 infection: reduction by 2year antiretroviral therapy. AIDS 24: 1733–1737.
- Brenchley JM, Price DA, Schacker TW, Asher TE, Silvestri G, et al. (2006) Microbial translocation is a cause of systemic immune activation in chronic HIV infection. Nat Med 12: 1365–1371.
- Martinez E, Larrousse M, Gatell JM (2009) Cardiovascular disease and HIV infection: host, virus, or drugs? Curr Opin Infect Dis 22: 28–34.

- Boccara F (2008) Cardiovascular complications and atherosclerotic manifestations in the HIV-infected population: type, incidence and associated risk factors. AIDS 22 Suppl 3: S19–26.
- Eden A, Price RW, Spudich S, Fuchs D, Hagberg L, et al. (2007) Immune activation of the central nervous system is still present after >4 years of effective highly active antiretroviral therapy. J Infect Dis 196: 1779–1783.
- Lichtfuss GF, Hoy J, Rajasuriar R, Kramski M, Crowe SM, et al. (2011) Biomarkers of immune dysfunction following combination antiretroviral therapy for HIV infection. Biomark Med 5: 171–186.
- Abdurahman S, Barqasho B, Nowak P, Cuong DD, Amogne W, et al. (2011) Elevated Flagellin-specific Antibodies during HIV-1 Infection. Abstract CROI 2011.
- Pelsers MM, Namiot Z, Kisielewski W, Namiot A, Januszkiewicz M, et al. (2003) Intestinal-type and liver-type fatty acid-binding protein in the intestine. Tissue distribution and clinical utility. Clin Biochem 36: 529–535.
- Sandler NG, Wand H, Roque A, Law M, Nason MC, et al. (2011) Plasma levels of soluble CD14 independently predict mortality in HIV infection. J Infect Dis 203: 780–790.
- Yukl SA, Gianella S, Sinclair E, Epling L, Li Q, et al. (2010) Differences in HIV burden and immune activation within the gut of HIV-positive patients receiving suppressive antiretroviral therapy. J Infect Dis 202: 1553–1561.
- Chege D, Sheth PM, Kain T, Kim CJ, Kovacs C, et al. (2011) Sigmoid Th17 populations, the HIV latent reservoir, and microbial translocation in men on long-term antiretroviral therapy. AIDS 25: 741–749.
- Bonora S, Nicastri E, Calcagno A, Gonzalez de Requena D, D'Ettorre G, et al. (2009) Ultrasensitive assessment of residual HIV viraemia in HAART-treated patients with persistently undetectable plasma HIV-RNA: a cross-sectional evaluation. J Med Virol 81: 400–405.
- Josephson F, Andersson MC, Flamholc L, Gisslen M, Hagberg L, et al. (2010) The relation between treatment outcome and efavirenz, atazanavir or lopinavir exposure in the NORTHIV trial of treatment-naive HIV-1 infected patients. Eur. J Clin Pharmacol 66: 349–357.
- Eden A, Andersson LM, Andersson O, Flamholc L, Josephson F, et al. (2010) Differential effects of efavirenz, lopinavir/r, and atazanavir/r on the initial viral decay rate in treatment naive HIV-1-infected patients. AIDS Res Hum Retroviruses 26: 533–540.
- Funaoka H, Kanda T, Kajiura S, Ohkaru Y, Fujii H (2011) Development of a high-specificity sandwich ELISA system for the quantification of human intestinal fatty acid-binding protein (I-FABP) concentrations. Immunol Invest 40: 223–242.
- Nowak P, Abdurahman S, Lindkvist A, Troseid M, Sonnerborg A (2012) Impact of HMGB1/TLR Ligand Complexes on HIV-1 Replication: Possible Role for Flagellin during HIV-1 Infection. Int J Microbiol 2012: 263836.
- Mizel SB, Bates JT (2010) Flagellin as an adjuvant: cellular mechanisms and potential. J Immunol 185: 5677–5682.
- Cassol E, Malfeld S, Mahasha P, van der Merwe S, Cassol S, et al. (2010) Persistent microbial translocation and immune activation in HIV-1-infected South Africans receiving combination antiretroviral therapy. J Infect Dis 202: 723-733.
- Wallet MA, Rodriguez CA, Yin L, Saporta S, Chinratanapisit S, et al. (2010) Microbial translocation induces persistent macrophage activation unrelated to HIV-1 levels or T-cell activation following therapy. AIDS 24: 1281–1290.
- Merlini E, Bai F, Bellistri GM, Tincati C, d'Arminio Monforte A, et al. (2011) Evidence for polymicrobic flora translocating in peripheral blood of HIVinfected patients with poor immune response to antiretroviral therapy. PLoS One 6: e18580.
- Marchetti G, Merlini E, Sinigaglia E, Iannotti N, Bai F, et al. (2012) Immune Reconstitution in HIV+ Subjects on Lopinavir/Ritonavir-Based HAART According to the Severity of Pre-Therapy CD4+. Curr HIV Res.
- Rajasuriar R, Booth D, Solomon A, Chua K, Spelman T, et al. (2010) Biological determinants of immune reconstitution in HIV-infected patients receiving

antiretroviral therapy: the role of interleukin 7 and interleukin 7 receptor alpha and microbial translocation. J Infect Dis 202: 1254–1264.

- Marchetti G, Cozzi-Lepri A, Merlini E, Bellistri GM, Castagna A, et al. (2011) Microbial translocation predicts disease progression of HIV-infected antiretroviral-naive patients with high CD4+ cell count. AIDS 25: 1385–1394.
- Douek DC, Roederer M, Koup RA (2009) Emerging concepts in the immunopathogenesis of AIDS. Annu Rev Med 60: 471–484.
- Anas A, van der Poll T, de Vos AF (2010) Role of CD14 in lung inflammation and infection. Crit Care 14: 209.
- Ayaslioglu E, Kalpaklioglu F, Kavut AB, Erturk A, Capan N, et al. (2012) The role of CD14 gene promoter polymorphism in tuberculosis susceptibility. J Microbiol Immunol Infect.
- Kanda T, Tsukahara A, Ueki K, Sakai Y, Tani T, et al. (2011) Diagnosis of ischemic small bowel disease by measurement of serum intestinal fatty acidbinding protein in patients with acute abdomen: a multicenter, observer-blinded validation study. J Gastroenterol 46: 492–500.
- Palmisano L, Giuliano M, Nicastri E, Pirillo MF, Andreotti M, et al. (2005) Residual viraemia in subjects with chronic HIV infection and viral load <50 copies/ml: the impact of highly active antiretroviral therapy. AIDS 19: 1843– 1847.
- Nicastri E, Palmisano L, Sarmati L, D'Ettorre G, Parisi S, et al. (2008) HIV-1 residual viremia and proviral DNA in patients with suppressed plasma viral load (<400 HIV-RNA cp/ml) during different antiretroviral regimens. Curr HIV Res 6: 261–266.
- Derikx JP, Vreugdenhil AC, Van den Neucker AM, Grootjans J, van Bijnen AA, et al. (2009) A pilot study on the noninvasive evaluation of intestinal damage in celiac disease using I-FABP and L-FABP. J Clin Gastroenterol 43: 727–733.
- Wiercinska-Drapalo A, Jaroszewicz J, Siwak E, Pogorzelska J, Prokopowicz D (2008) Intestinal fatty acid binding protein (I-FABP) as a possible biomarker of ileitis in patients with ulcerative colitis. Regul Pept 147: 25–28.
- Pilon AA, Lum JJ, Sanchez-Dardon J, Phenix BN, Douglas R, et al. (2002) Induction of apoptosis by a nonnucleoside human immunodeficiency virus type 1 reverse transcriptase inhibitor. Antimicrob Agents Chemother 46: 2687–2691.
- Blas-Garcia A, Esplugues JV, Apostolova N (2011) Twenty years of HIV-1 nonnucleoside reverse transcriptase inhibitors: time to reevaluate their toxicity. Curr Med Chem 18: 2186–2195.
- Rizza SA, Badley AD (2008) HIV protease inhibitors impact on apoptosis. Med Chem 4: 75–79.
- Badley AD (2005) In vitro and in vivo effects of HIV protease inhibitors on apoptosis. Cell Death Differ 12 Suppl 1: 924–931.
- Lodes MJ, Cong Y, Elson CO, Mohamath R, Landers CJ, et al. (2004) Bacterial flagellin is a dominant antigen in Crohn disease. J Clin Invest 113: 1296–1306.
- Lien E, Aukrust P, Sundan A, Muller F, Froland SS, et al. (1998) Elevated levels of serum-soluble CD14 in human immunodeficiency virus type 1 (HIV-1) infection: correlation to disease progression and clinical events. Blood 92: 2084– 2092.
- Jakobsson HE, Jernberg C, Andersson AF, Sjolund-Karlsson M, Jansson JK, et al. (2010) Short-term antibiotic treatment has differing long-term impacts on the human throat and gut microbiome. PLoS One 5: e9836.
- Jernberg C, Lofmark S, Edlund C, Jansson JK (2010) Long-term impacts of antibiotic exposure on the human intestinal microbiota. Microbiology 156: 3216–3223.
- Mermin J, Lule J, Ekwaru JP, Malamba S, Downing R, et al. (2004) Effect of cotrimoxazole prophylaxis on morbidity, mortality, CD4-cell count, and viral load in HIV infection in rural Uganda. Lancet 364: 1428–1434.
- Campbell JH, Burdo TH, Autissier P, Bombardier JP, Westmoreland SV, et al. (2011) Minocycline inhibition of monocyte activation correlates with neuronal protection in SIV neuroAIDS. PLoS One 6: e18688.
- Kalambokis GN, Tsianos EV (2012) Rifaximin reduces endotoxemia and improves liver function and disease severity in patients with decompensated cirrhosis. Hepatology 55: 655–656.